

## AGENDA

### **PacMOSSI short course: Insecticide Resistance, Product Efficacy and Mosquito Identification**

Venue: QIMR Berghofer Medical Research Institute,  
Brisbane, Queensland, Australia.

Dates: Session 1: 26<sup>th</sup> to 30<sup>th</sup> September 2022  
Session 2: 10<sup>th</sup> to 14<sup>th</sup> October 2022

#### ***Participants:***

Osiro Lorin (Palau)  
Oni Lewakulati (Fiji)  
Siaola Mahe (Tonga)  
Paulo Pemitia Seuseu (Samoa)  
Boni Sebayang (JCU, Australia)  
Bram Van de Straat (JCU, Australia)  
Lekon Tagavi (Vanuatu)  
Nooroa Tuakeu (Cook Islands)  
Charlie Iro-ofa (Solomon Islands)  
Lolo Viamoana Tuumuli (Tokelau)  
Tamera Shimuzu (Guam)  
Felila Peter (Nauru)  
Metotagivale Pitoitua (American Samoa)

#### ***Demonstrators / mentors / observers***

Hasan Al Amin (QIMRB)  
Tom Burkot (JCU)  
Greg Devine (QIMRB)  
Narayan Gyawali (QIMRB)  
Brian Johnson (QIMRB)  
Tessa Knox (WHO)  
Amanda Murphy (WHO)  
Tanya Russell (JCU)  
Sala Saketa (SPC)  
Andrew van den Hurk (QH)

## Detailed schedule:

*Venues: Level 3 training room (L3) or suite 2, level 13 insectary (S2).*

Day	Time	Programme	Place	Trainers
Monday	9:00 to 9:15	Receive trainees at reception		Narayan Gyawali (NG) Greg Devine (GD)
	9:15 to 9:50	Welcome, introductions. QIMRB induction (canteen, fire drill etc) and quarantine rules.	L3	Tom Burkot (TB) Tanya Russell (TR) NG, GD
	10:00 to 11:00	Theory: Basics of mosquito identification (focus on larvae)	L3	Brian Johnson (BJ) NG
	11:05 to 12:00	Theory: Identifying adult specimens to species	L3	Andrew van den Hurk (AvdH)
	12:00 to 13:00	Lunch		
	13:00 to 15:00	Demonstration: Rearing mosquitoes: Egg hatching Feeding Larvae Pupae collection ("picking") Adult cage preparation Blood feeding Oviposition substrates	S2	NG, GD Hassan Al Amin (HMA)
	15:00 to 15:30	Break		
	15:30 to 17:00	<b>Practical:</b> Mosquito Identification: mosquito or not? Differentiating male and female mosquitoes.	L3	AvdH, HMA, NG, BJ
Tuesday	9:00 to 9:45	Theory: Introduction to insecticide resistance and assay methods (tube tests and bottle assays)	L3	GD
	9:50 to 10:45	Theory: Efficacy testing I (LLIN/IRS, Cone test)	L3	GD
	10:45 to 11:00	Break		
	11:00 to 12:00	Theory: Efficacy testing II (Tunnel Test, Space spray)	L3	GD, TR, TB
	12:00 to 13:00	Lunch		

	13:00 to 13:30	Practical: making an insecticide dilution	S2	NG
	13:30 to 15:00	CDC Bottle assays on resistant and susceptible mosquitoes	S2	HMA, NG, BJ
	15:00 to 15:30	Break		
	15:30 to 17:00	WHO tube tests on resistant and susceptible mosquitoes	S2	HMA, NG, BJ
Wednesday	9:00 to 09:30	Theory: larval bioassay	L3	NG
	9:30 to 11:00	Practical: larval bioassay	S2	NG, HMA
	11:00 to 11:10	Break		
	11:10 to 13:00	Practical: Recording and reporting results from the bottle and tube assays	L3	NG, HMA
	13:00 to 14:00	Lunch		
	14:00 onward	Field visit: Mosquito trap set	Field	BJ, NG
Thursday	9:00 to 11:00	Practical: Mosquito identification from local traps (including those that participants may have brought with them).	S2	AvdH, BJ
	11:00 to 11:10	Break		
	11:10 to 12:00	Practical: Recording and reporting the results from larval assays.	L3	NG
	12:00 to 13:00	Lunch		
	13:00 to 16:00	Practical: Cone tests for monitoring product efficacy	S2	HMA, GD, NG
<b>COURSE DINNER</b>				
Friday 30 <sup>th</sup>	9:00 to 10:00	Recording and reporting results from cone tests	L3	HMA, NG
	10:00-10:10	Break		
	10:00 to 11:30	Practical: Species identification, introduction to course supplements	L3	NG, BJ
	11:30 to 12:30	Course review and questions Farewell	L3	NG, GD, TR, TB
	12:30 -	Lunch/Close		