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A guide to mosquitoes in the Pacific 2023

Authors: Tanya L. Russell and Thomas R. Burkot





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Pacific Mosquito Surveillance
Strengthening for Impact

A guide to mosquitoes in the Pacific

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Authors: Tanya L. Russell and Thomas R. Burkot

Affiliation: Australian Institute of Tropical Health and Medicine, James Cook University, Cairns,
Australia

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Dedication

Dedicated to all scientists who have devoted their career to studying the biology, taxonomy and distribution of mosquitoes in the Pacific. In particular, John Belkin, whose monumental work "Mosquitoes of the South Pacific" published 60 years ago remains one of the most influential publications in this field. Belkin served as the Commanding Officer of the 420th Malaria Survey Detachment for 21 months in the Solomon Islands (1943–1945), and it was during this time that he conducted extensive and meticulous mosquito collections that were the basis for his work. Brian Taylor, while based in the Solomon Islands (1969–1973), discovered the phenomenon of behavioural resistance in *Anopheles farauti*. Elizabeth Marks described the taxonomy of a number of mosquitoes, most notably *Aedes polynesiensis*. Joan Bryan conducted seminal research detailing the speciation of the *Anopheles punctulatus* complex. There are many others who have contributed to the field including Margaret Spencer, Wallace Peters, Dirk Metselaar, Kenneth Knight, Richard Bohart, George Bohart, Marshall Laird, Mandayam Iyengar, Peter Mattingly, Mario Maffi, Harry Hoogstraal, Rudolf Slooff and Harry Standfast. Especially, we acknowledge and dedicate this book to the residents of the thousands of islands in the Pacific region who supported the work of everyone who worked on mosquitoes in the Pacific by welcoming us to their villages and sharing their food and knowledge.

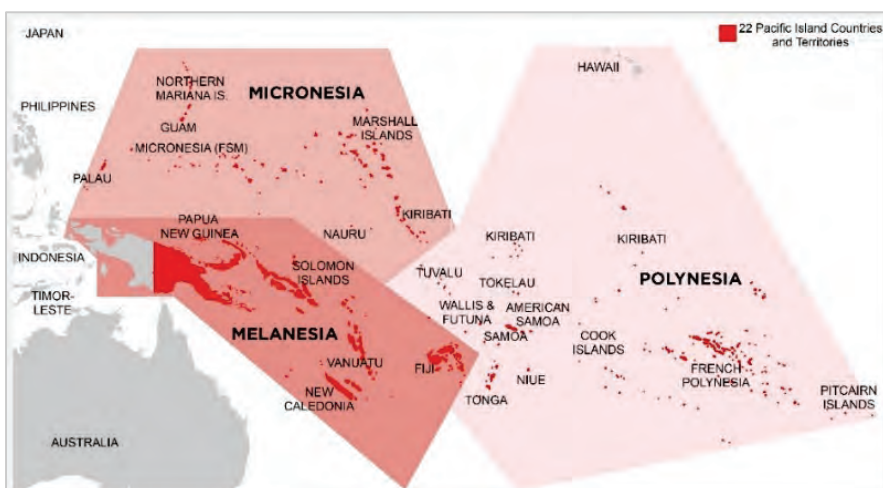
Introduction

The Pacific is home to 11.4 million people residing in 22 countries and territories across the region. There are thousands of islands in the Pacific Ocean, which are divided into three main groups known as Micronesia, Melanesia and Polynesia. Pacific Islands are composed of four basic types: continental islands, high islands, coral reefs and uplifted coral platforms. This unique geography supports highly diverse ecosystems, home to 423 known mosquito species, some of which transmit arboviruses (e.g. dengue, Zika, chikungunya and Ross River, Japanese encephalitis, amongst others), human malaria parasites (e.g. *Plasmodium falciparum*, *P. vivax*, *P. ovale* and *P. malariae*) and lymphatic filariasis (*Wuchereria bancrofti*).

Mosquitoes are considered to be the most dangerous animals in the world, due to their ability to transmit numerous pathogens. Recently, the Pacific has been experiencing unprecedented outbreaks of dengue, chikungunya and Zika virus¹ alongside ongoing malaria² and lymphatic filariasis³ transmission with direct effects on morbidity and mortality. Further, there is likely ongoing Ross River virus circulation in many Pacific countries^{4,5} and Japanese encephalitis is endemic in Papua New Guinea^{6,7}. The ongoing transmission of mosquito-borne diseases places a heavy toll on already fragile health systems with local economic and social repercussions.

Vector control interventions have one of the highest returns on investment in public health,⁸ and are the primary method for controlling malaria⁹ and *Aedes*-borne arboviruses¹⁰. While vector control is secondary to mass drug administration in campaigns to eliminate lymphatic filariasis, it is an effective tool that could accelerate elimination programs and prevent resurgence¹¹⁻¹³. Integrated and stratified vector and disease programs seek to ensure the limited human and financial resources simultaneously prevent and control both the mosquitoes and the pathogens. The foundation of successful integrated and stratified programs is knowledge of the distributions and behaviours of the vectors. Such programs require strong vector surveillance programs because the behaviours or the vectors in the Pacific are dynamic and can change rapidly in response to the selective pressures exerted by vector control.

This handbook provides practical and basic biological information on the behaviours and distribution of the mosquitoes of the Pacific region as they are presently known. This handbook is a foundational reference for all those working towards reducing the transmission of mosquito-borne diseases in the region. An overview of the diversity and distribution of mosquitoes throughout the Pacific, including checklists for the species present in each of the 22 Pacific Island countries and territories, is provided. For the mosquitoes that are common and/or of medical importance, one-page profiles are provided that include information about their key behaviours. While much is known about some key vectors, surprisingly little is known about many others. The dynamic capacity of vectors to invade new areas or adapt their behaviours to changing environments ensures that while every effort has been made to ensure the information contained herein is accurate at this time, it may not be in the future.



A guide to mosquitoes in the Pacific

This handbook includes bionomic summaries of 42 disease vectors and/or those most commonly encountered (either at the species or complex level). The mosquito species are presented in alphabetical order and information on the distribution, aquatic habitats and adult female behaviours are summarised. The compilation of information regarding the distribution of mosquito species is based on published literature and personal communications (up to and including year 2021). Where rigorous and extensive vector surveillance hasn't detected a species since 2010, it has been listed as eliminated. However, it should be noted that the absence of a mosquito species is difficult to confirm and that distributions change over time.

The geography of the Pacific has fostered an array of unique mosquito fauna whose distributions range from spanning the Pacific to being limited to single islands. The mosquito fauna of the Pacific is unique, highly endemic and understudied. The endemism of the Pacific mosquito fauna is interesting and significant. There is a strong differentiation in species across the islands of the entire region, with 72% (n = 299) of species only found from a single country, and often on individual isolated islands.

In the Southwest Pacific there are over 42 species of *Anopheles*, which are found in the islands of New Guinea, the Solomon Islands and Vanuatu. The region supports the most intense malaria transmission outside Africa. The malaria transmission system is complex and diverse with at least 36 species found in Papua New Guinea and only a single vector found in Vanuatu. The Pacific remains the one region in the world where physiological resistance to pyrethroid-based insecticides is not common in the dominant *Anopheles* species. In contrast, the primary vector, *Anopheles farauti s.s.*, has markedly changed its behaviour to avoid contact with insecticides, a phenomenon that was originally detected in the Solomon Islands¹⁴ and later in Papua New Guinea¹⁵, and known as behavioural resistance.¹⁶ Incursions of *Anopheles* into new areas has occurred, including from species that naturally occur west of the Weber Line into the region, and also from Pacific species into new areas. As a result, anophelism without malaria occurs in New Caledonia and Micronesia.^{17,18}

The genus *Aedes* is the largest in the tribe Aedini and includes many vectors of debilitating viral diseases of humans. There are 119 *Aedes* species in the Pacific. The most complex array of *Aedes* arbovirus vectors in the world is found in the Pacific region, where at least 12 species of the *Stegomyia* subgenus are dengue vectors. Both *Aedes aegypti* and *Aedes albopictus* have invaded the region with ever increasing distributions. While *Ae. aegypti* and *Ae. albopictus* are the primary vectors, there have been dengue and chikungunya outbreaks on islands when these species were absent.^{19,20}

The genus *Culex* is represented by 113 different species in the Pacific. Some of these species are vectors of arboviruses (e.g. Ross River virus, Japanese encephalitis virus) and lymphatic filariasis. Some *Culex* species are extremely widespread across the globe, while others are endemic to the Pacific; for example, *Cx. roseni* in French Polynesia, *Cx. samoensis* in the Samoan islands, *Cx. pacificus* in Vanuatu and *Cx. iyengari* in New Caledonia.

Medical importance

In the Pacific, malaria transmission is endemic in Papua New Guinea, the Solomon Islands and Vanuatu. *Aedes*-borne arboviruses, including dengue, Zika and chikungunya, occur across the Pacific. Lymphatic filariasis has long been endemic across the Pacific where it is caused by the species *Wuchereria bancrofti*. Further, there is likely ongoing Ross River virus circulation in many Pacific countries, and Japanese encephalitis is endemic in Papua New Guinea. It is important to note that different mosquito species are responsible for transmitting different pathogens between animal hosts. However, not all mosquito species are capable of pathogen transmission due to differences in their biology and ecology and the ability of the pathogen to multiply or develop in the mosquito. Further, vectorial capacity can vary greatly between populations, and a species may not act as a vector in all localities where it is present.

The vectors are often referred to as either primary or secondary vectors. Primary vectors are the species that are responsible for the majority of the transmission of a pathogen (they can become infectious, are abundant and readily bite humans). Secondary vectors are species that are competent but have a more limited role in disease transmission (they can become infectious, they may be less abundant in the landscape, or feed largely on non-human hosts). The following tables list the species demonstrated to be competent vectors in the Pacific. Notes regarding the primary and secondary vector status of species is included only in the species profiles.

Vectors of malaria in the Pacific

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species			
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Anopheles</i>	<i>bancroftii</i>	<i>bancroftii</i>	<i>bancroftii</i> A			
						<i>bancroftii</i> B			
						<i>bancroftii</i> C			
						<i>bancroftii</i> D			
				<i>barbirostris</i>	<i>barbirostris</i>	<i>barbirostris</i>			
						<i>campestris</i>			
						<i>hyrcanus</i>			
				<i>Cellia</i>	-	-	<i>indefinitus</i>		
							<i>karwari</i>		
							<i>litoralis</i>		
			<i>vagus</i>						
			<i>longirostris</i>				-	-	<i>longirostris</i> A
									<i>longirostris</i> B
				<i>longirostris</i> C1					
				<i>longirostris</i> C2					
				<i>longirostris</i> D					
				<i>longirostris</i> E					
				<i>longirostris</i> F					
				<i>longirostris</i> G					
			<i>punctulatus</i>	<i>farauti</i>	<i>farauti</i>	<i>subpictus</i>			
<i>subpictus</i>									
<i>punctulatus</i>									
<i>farauti</i> 4									
<i>farauti</i> 8									
<i>farauti</i> s.s.									
<i>koliensis</i>	<i>koliensis</i>	<i>koliensis</i>	<i>hinesorum</i>						
			<i>oreios</i>						
						<i>koliensis</i> 1			
						<i>koliensis</i> 3			

Other suspected (but not confirmed) vectors are *Anopheles hilli*, *Anopheles lungae* s.s., *Anopheles meraukensis*, *Anopheles novaguinensis* and *Anopheles solomonis*. Note that the vector status of each species comprising the complexes *Anopheles bancroftii* s.l., *Anopheles longirostris* s.l. and *Anopheles koliensis* s.l. has not been confirmed.

Vectors of dengue in the Pacific

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
				<i>aegypti</i>	-	<i>aegypti</i>
						<i>albopictus</i>
						<i>cooki</i>
						<i>hebrideus</i>
						<i>hensilli</i>
						<i>kesseli</i>
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>marshallensis</i>
						<i>polynesiensis</i>
						<i>pseudoscutellaris</i>
						<i>rotumae</i>
						<i>scutellaris</i>
					tongae	<i>tabu</i>
						<i>tongae s.s.</i>

Vectors of chikungunya in the Pacific

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
				<i>aegypti</i>	-	<i>aegypti</i>
						<i>albopictus</i>
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>hensilli</i>
						<i>polynesiensis</i>

Note that it is unknown if other dengue vectors are involved in chikungunya transmission.

Vectors of Zika in the Pacific

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
				<i>aegypti</i>	-	<i>aegypti</i>
						<i>albopictus</i>
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>hensilli</i>
						<i>polynesiensis</i>

Note that it is unknown if other dengue vectors are involved in Zika transmission.

Vectors of lymphatic filariasis in the Pacific

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species		
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Anopheles</i>	<i>bancroftii</i>	<i>bancroftii</i>	<i>bancroftii</i> A		
						<i>bancroftii</i> B		
						<i>bancroftii</i> C		
						<i>bancroftii</i> D		
				<i>barbirostris</i>	<i>barbirostris</i>	<i>barbirostris</i>		
			-	<i>subpictus</i>	<i>subpictus</i>			
			<i>Cellia</i>	<i>punctulatus</i>	-	<i>punctulatus</i>		
					<i>farauti</i>	<i>farauti</i> s.s.		
					<i>koliensis</i>	<i>hinesorum</i>		
					-	<i>koliensis</i> 1		
-	<i>koliensis</i> 3							
Culicinae	Aedini	<i>Aedes</i>	<i>Finlaya</i>	<i>kochi</i>	-	<i>oceanicus</i>		
					-	<i>fijiensis</i>		
					-	<i>kochi</i>		
					-	<i>samoanus</i>		
				<i>Ochlerotatus</i>	<i>emiphals</i>	-	<i>vigilax</i>	
			<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>cooki</i>		
					-	<i>kesseli</i>		
					-	<i>marshallensis</i>		
					-	<i>polynesiensis</i>		
					-	<i>pseudoscutellaris</i>		
Culicini	<i>Culex</i>	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i>		
						<i>sitiens</i>	-	<i>annulirostris</i>
						<i>Oculeomyia</i>	-	<i>bitaeniorhynchus</i>
						<i>bitaeniorhynchus</i>	-	<i>bitaeniorhynchus</i>
						<i>uniformis</i>	-	<i>uniformis</i>

Note that the vector status of each species comprising the complexes *Anopheles bancroftii* s.l. and *Anopheles koliensis* s.l. has not been confirmed. A suspected (but not confirmed) vector is *Aedes tutuila*.²¹

Vectors of Ross River virus in the Pacific

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species						
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Anopheles</i>	<i>bancroftii</i>	<i>bancroftii</i>	<i>bancroftii</i> A						
			<i>Cellia</i>	<i>annulipes</i>	<i>annulipes</i>	<i>annulipes</i>						
			<i>Edwardsaedes</i>	-	-	<i>imprimens</i>						
			<i>Finlaya</i>	<i>kochi</i>	-	<i>kochi</i>						
		Aedini	<i>Aedes</i>	<i>Ochlerotatus</i>	-	-	<i>normanensis</i>					
					<i>emiphals</i>	-	<i>vigilax</i>					
				<i>Rampamyia</i>	<i>notoscriptus</i>	-	<i>notoscriptus</i>					
					<i>aegypti</i>	-	<i>aegypti</i>					
				<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>albopictus</i>					
						-	<i>polynesiensis</i>					
Culicinae		<i>Verrallina</i>	<i>Verrallina</i>	-	-	<i>carmenti</i>						
						<i>funerea</i>						
						<i>lineata</i>						
						<i>annulirostris</i>						
						Culicini	<i>Culex</i>	<i>Culex</i>	<i>sitiens</i>	-	-	<i>gelidus</i>
												<i>palpalis</i>
												<i>linealis</i>
						Mansoniini	<i>Mansonia</i>	<i>Mansonioides</i>	-	-	-	<i>septempunctata</i>
												<i>uniformis</i>

A suspected (but not confirmed) vector is *Anopheles hinesorum* in Papua New Guinea.²²

Vectors of Japanese encephalitis virus in the Pacific

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species						
Culicinae						<i>Aedimorphus</i>						
						<i>vexans</i>						
						<i>Aedes</i>	<i>Ochlerotatus</i>	<i>emiphals</i>	-	<i>vigilax</i>		
							<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>albopictus</i>		
						<i>Armigeres</i>	<i>Armigeres</i>	-	-	<i>subalbatus</i>		
								<i>pipiens</i>	-	<i>fuscocephala</i>		
									<i>pipiens</i>	<i>quinquefasciatus</i>		
						Culicini	<i>Culex</i>	<i>Culex</i>	<i>sitiens</i>	-	-	<i>annulirostris</i>
												<i>gelidus</i>
												<i>palpalis</i>
						<i>sitiens</i>						
		<i>Oculeomyia</i>	-	<i>bitaeniorhynchus</i>	<i>bitaeniorhynchus</i>	<i>bitaeniorhynchus</i>						

Note that in the Pacific, Japanese encephalitis is endemic in Papua New Guinea^{6,7} and was recently introduced to mainland Australia.²³

List of mosquitoes species or complexes included

Anopheles bancroftii s.l. Giles 1902
Anopheles farauti s.s. Laveran 1902
Anopheles farauti 4 (unnamed)
Anopheles farauti 8 (unnamed)
Anopheles hinesorum Schmidt 2001
Anopheles karwari (James) 1903
Anopheles koliensis s.l. Owen 1945
Anopheles longirostris s.l. Brug 1928
Anopheles oreios Bangs & Harbach 2015
Anopheles punctulatus Dönitz 1901
Aedes aegypti (Linnaeus) 1762
Aedes albolineatus (Theobald) 1904
Aedes albopictus (Macquart) 1903
Aedes cooki Belkin 1962
Aedes fijiensis Marks 1947
Aedes guamensis Farner & Bohart 1944
Aedes hebrideus Edwards 1962
Aedes hensilli Farner 1945
Aedes horrescens Edwards 1935
Aedes kesseli Huang & Hitchcock 1980
Aedes kochi (Donitz) 1901
Aedes marshallensis (Stone & Bohart) 1944
Aedes notoscriptus (Skuse) 1889
Aedes oceanicus Belkin 1962
Aedes polynesiensis Marks 1951
Aedes pseudoscutellaris (Theobald) 1901
Aedes rotumae Belkin 1962
Aedes samoanus (Grünberg) 1913
Aedes scutellaris (Walker) 1858
Aedes tabu Ramalingam & Belkin 1965
Aedes tongae s.s. Edwards 1926
Aedes tutuilae Ramalingam & Belkin 1965
Aedes upolensis Marks 1957
Aedes vexans (Meigen) 1830
Aedes vigilax (Skuse) 1889
Culex annulirostris Skuse 1889
Culex bitaeniorhynchus Giles 1901
Culex gelidus Theobald 1901
Culex quinquefasciatus Say 1823
Culex sitiens Wiedemann 1828
Culex tritaeniorhynchus Giles 1901
Mansonia uniformis (Theobald) 1901



Aedes aegypti

Image source: CDC Public Health Image Library



Anopheles farauti



Culex quinquefasciatus

Anopheles bancroftii s.l. Giles 1902

Distribution of *Anopheles bancroftii* s.l. throughout the Pacific Islands

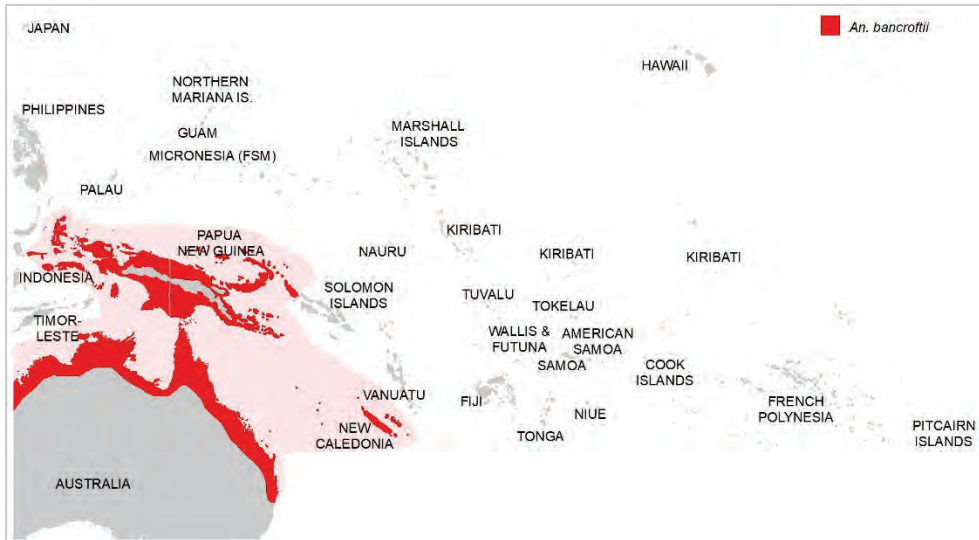


Photo source:
Stephen Doggett

Bionomics of *Anopheles bancroftii* s.l.

Subgenus	<i>Anopheles</i>
Distribution	A complex of four species found across Australia, West Papua, the Moluccas, and Papua New Guinea. ²⁴⁻³² <i>Anopheles bancroftii</i> A (ITS2 genotype A1) was introduced in New Caledonia in 2017. ¹⁷
Aquatic habitats	The aquatic habitats are mainly large vegetated freshwater swamps and wetlands. Larvae have also been collected from the margins of streams and drains. ^{32,33} The larvae prefer shaded areas and are commonly associated with surface aquatic plants (including water ferns and water hyacinth) or debris.
Feeding times	Feeds primarily at night-time (nocturnal), and will also feed during the day among well-shaded vegetation. ³²
Blood meal hosts	Opportunistically feeds on humans or other animals. The preference of the species to feed on humans differs among localities. ^{32,34,35}
Feeding location	Readily feeds both indoors and outdoors, with the preferred feeding location differing among localities. ³²
Resting habits	Rests outdoors in vegetation, and those that entered houses to feed will rest inside for a short period. ³²
Flight range	Long flight range of up to 4–5 km. Can fly over large bodies of water to reach the opposite bank. ³⁶
Vector status	<i>Plasmodium</i> spp. (secondary vector), ^{25,32,37,38} <i>Wuchereria bancrofti</i> (secondary vector as demonstrated in West Papua, Indonesia), ³⁶ Ross River virus and other zoonotic arboviruses. ^{32,39}

Anopheles farauti s.s. Laveran 1902

Distribution of *Anopheles farauti* s.s. throughout the Pacific Islands

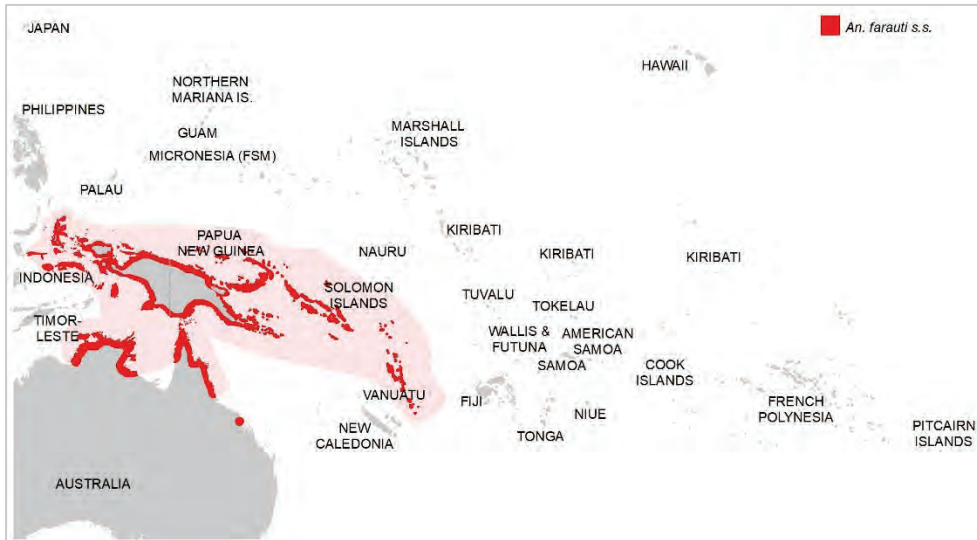


Photo source:
Stephen Doggett

Bionomics of *Anopheles farauti* s.s.

Subgenus	<i>Cellia</i>
Distribution	Occurs in the Moluccas, on New Guinea, in northern Australia, the Solomon Islands and Vanuatu within 2 km from the coast. ^{25,29,33,40-50} Formerly <i>Anopheles farauti</i> 1.
Aquatic habitats	Immatures typically occupy saline or brackish water habitats, including lagoons, coastal mangrove swamps and brackish pools. Habitats are often shaded and vegetated. Will occupy freshwater habitats. ^{48,51}
Feeding times	Feeds primarily at night (crepuscular and nocturnal). Peak biting activity varies by location: in the Solomon Islands peak biting is from 1830–2100, ⁵² and in Papua New Guinea and Vanuatu it is from 1900–0100. ^{15,53}
Blood meal hosts	Opportunistically feeds on humans or animals. ⁵⁴ The human blood index for the species varies greatly, often depending on the availability of vertebrate hosts. ^{35,55}
Feeding location	While the majority of feeding occurs outdoors, <i>An. farauti</i> s.s. will also readily enter houses for blood meals ⁵⁶ . The proportion of exophagy varies by location.
Resting habits	<i>An. farauti</i> s.s. will rest for short periods of time indoors after blood feeding indoors but this species is mostly exophilic, tending to rest mainly outdoors. ⁵⁷
Flight range	While the flight range of <i>An. farauti</i> s.s. is usually short (within 100 m), they have been recorded to fly up to 1 km to seek blood meals where hosts are not located near the aquatic habitats.
Vector status	<i>Plasmodium</i> spp. (primary vector), ^{15,25,32,45,53,58-61} <i>Wuchereria bancrofti</i> (primary vector) ⁶²⁻⁶⁶ and other zoonotic arboviruses. ²⁴

Anopheles farauti 4 (unnamed)

Distribution of Anopheles farauti 4 throughout the Pacific Islands

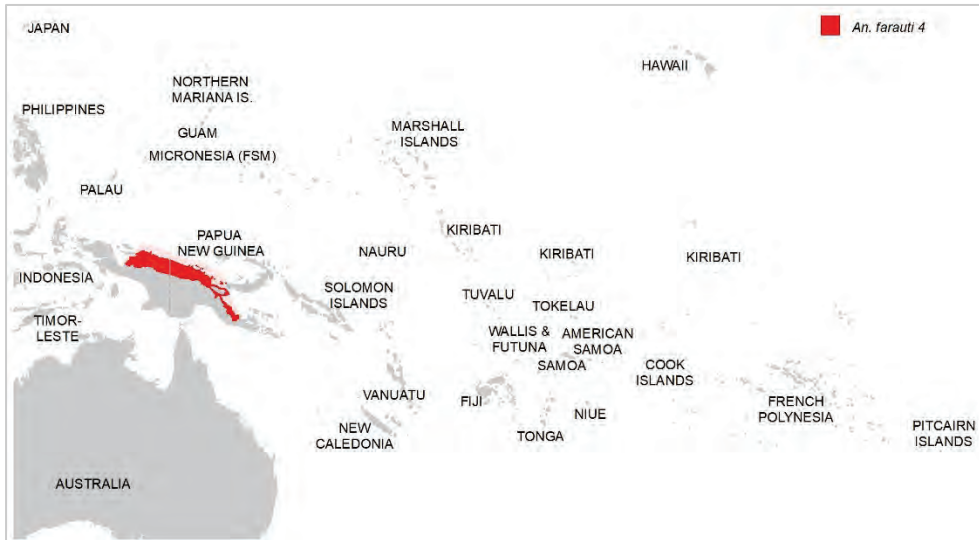


Photo source:
Stephen Doggett

Bionomics of Anopheles farauti 4

Subgenus	<i>Cellia</i>
Distribution	Found throughout the inland lowland river valleys and flood plains north of the central highlands on New Guinea island. ^{25,33,53,58,67,68}
Aquatic habitats	The aquatic habitats are mostly small transient sites exposed to sunlight with little vegetation, such as pools in rivers or streambeds, wheel ruts, puddles, hoof and footprints, pig wallows and shallow drains. ⁶⁷
Feeding times	Feeds throughout the night, but most of the feeding occurs early in the evening (crepuscular and nocturnal). ^{53,58,68}
Blood meal hosts	Highly anthropophilic with a strong preference to feed on humans. ^{35,53,58}
Feeding location	Readily feeds both indoors and outdoors. ⁵⁸
Resting habits	Unknown.
Flight range	Unknown.
Vector status	<i>Plasmodium</i> spp. (primary vector). ^{25,53,58}

Anopheles farauti 8 (unnamed)

Distribution of Anopheles farauti 8 throughout the Pacific Islands

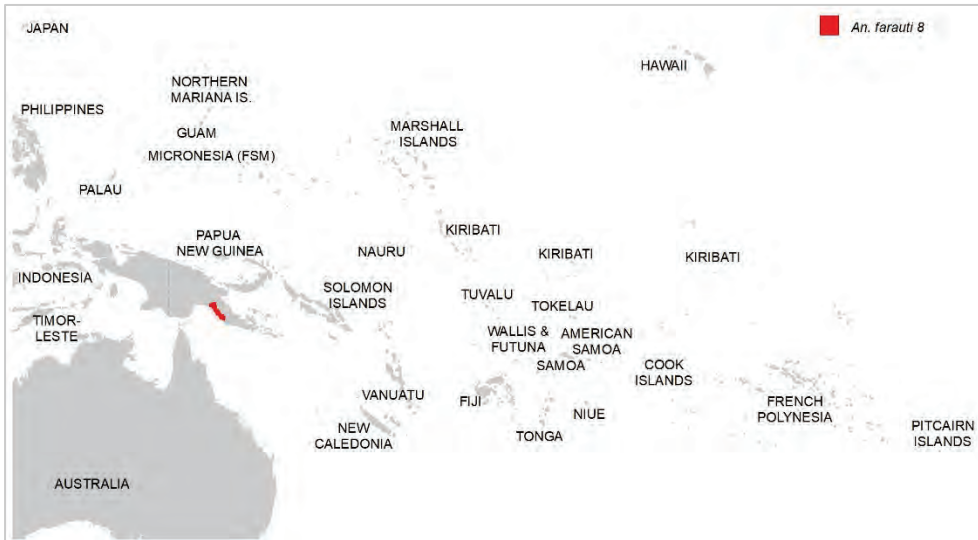


Photo source:
Stephen Doggett

Bionomics of Anopheles farauti 8

Subgenus	<i>Cellia</i>
Distribution	Has only been found in the inland lowland areas on the east side of the Gulf of Papua in Papua New Guinea. ^{33,67,69}
Aquatic habitats	Unknown.
Feeding times	Unknown.
Blood meal hosts	Unknown.
Feeding location	Unknown.
Resting habits	Unknown.
Flight range	Unknown.
Vector status	<i>Plasmodium</i> spp. (secondary vector). ²⁵

Anopheles hinesorum Schmidt 2001

Distribution of *Anopheles hinesorum* throughout the Pacific Islands

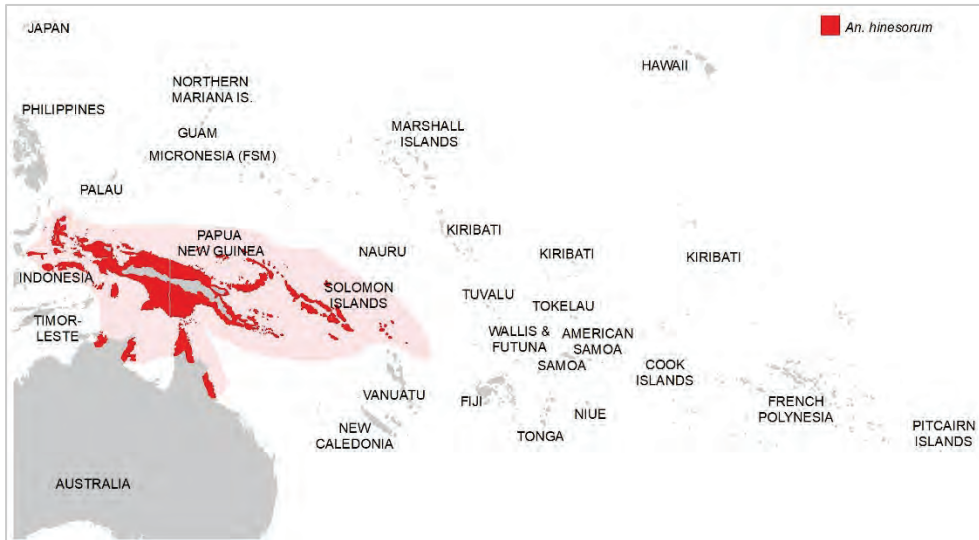


Photo source:
Stephen Doggett

Bionomics of *Anopheles hinesorum*

Subgenus	<i>Cellia</i>
Distribution	Is found near the coast, inland and on elevated areas. In Papua New Guinea this species is most frequently found in lowland inland river valleys and flood plains. ^{41,48-50,67,70-72} Formerly <i>Anopheles farauti</i> 2.
Aquatic habitats	Aquatic habitats include a range of water bodies, both natural – ground pools, swamps and the edges of streams and rivers – and human-made drains, ditches and wheel ruts as well as pig wallows. ^{33,48}
Feeding times	Feeds during the night, with feeding activity peaking early in the evening and then declining through the rest of the night (crepuscular and nocturnal). ^{53,73}
Blood meal hosts	Host feeding preference is extremely variable across its range. In Papua New Guinea, <i>An. hinesorum</i> will opportunistically feed on humans and mammals, ³⁵ while in the Solomon Islands it is predominantly zoophagic. ³³
Feeding location	Tendency to feed outdoors. ⁵³
Resting habits	Most likely rests outdoors, as inferred by an absence of <i>An. hinesorum</i> in indoor resting collections.
Flight range	Unknown
Vector status	<i>Plasmodium</i> spp. (primary vector in New Guinea island, non-vector in the Solomon Islands except in Western Province where it is a secondary vector), ^{25,49} <i>Wuchereria bancrofti</i> (primary vector in New Guinea). ⁶² A potential vector of Ross River virus in Papua New Guinea. ²²

Anopheles karwari (James 1903)

Distribution of *Anopheles karwari* throughout the Pacific Islands



Photo source: The Government of the Hong Kong Special Administrative Region

Bionomics of *Anopheles karwari*

Subgenus	<i>Cellia</i>
Distribution	Found north of the central highlands on New Guinea island, as well as in southeast Asia. ^{27,28,32,74} A rare species in Papua New Guinea, Indonesia and the Philippines.
Aquatic habitats	Found in natural and man-made shaded habitats containing freshwater, such as marshes, slow-moving streams, seepages, ground and rock pools, springs and irrigation channels. ²⁸
Feeding times	Unknown.
Blood meal hosts	Unknown.
Feeding location	Unknown.
Resting habits	Unknown.
Flight range	Unknown.
Vector status	<i>Plasmodium</i> spp. (secondary vector). ^{25,32,75}

Anopheles koliensis s.l. Owen 1945

Distribution of *Anopheles koliensis* throughout the Pacific Islands

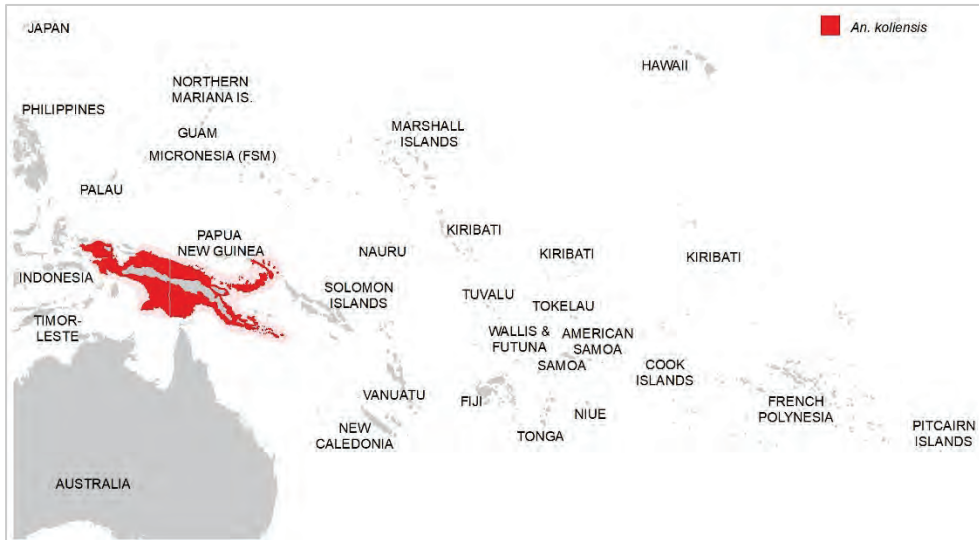


Photo source: Walter Reed Biosystematics Unit

Bionomics of *Anopheles koliensis* s.l.

Subgenus	<i>Cellia</i>
Distribution	This complex is found throughout New Guinea, New Britain and New Ireland Islands, ^{27,28,30,33,53,67,74,76,77} and is potentially 2–3 cryptic species. ⁷⁸ In the Solomon Islands, it is thought to have been eliminated. ¹⁶ It is mostly an inland species usually below 300 m.
Aquatic habitats	The main aquatic habitats are wheel tracks, drains, natural ground pools and freshwater swamps. ⁶⁷
Feeding times	Feeds during the night, with peak activity late at night between 2200–0200 (nocturnal). ⁵³
Blood meal hosts	Highly anthropophilic with a strong preference to feed on humans, but will also feed on pigs and dogs. ^{35,55}
Feeding location	Readily feeds both indoors and outdoors; endophagy varies by location.
Resting habits	May rest indoors for a short time after feeding indoors; most resting is outdoors.
Flight range	Unknown.
Vector status	<i>Plasmodium</i> spp. (primary vector), ^{15,25,53,75,79} <i>Wuchereria bancrofti</i> (primary vector). ^{32,80}

Anopheles longirostris s.l. Brug 1928

Distribution of *Anopheles longirostris* throughout the Pacific Islands

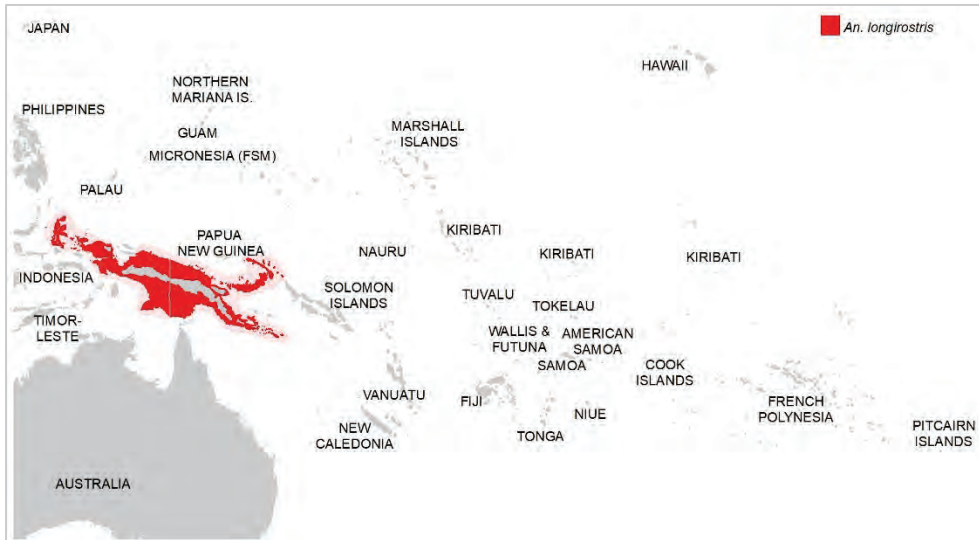


Photo source:
Walter Reed
Biosystematics Unit

Bionomics of *Anopheles longirostris* s.l.

Subgenus	<i>Cellia</i>
Distribution	Widespread throughout the coastal and inland lowland areas of New Guinea island and the Moluccas. ^{27,38,74,81,82} Occurs in the jungle or near its fringe and appears to have a limited association with humans. ³⁸ A complex of nine cryptic species. ⁸²
Aquatic habitats	Prefers specific aquatic habitats: pools located in jungle or densely vegetated areas. ^{25,38}
Feeding times	Unknown.
Blood meal hosts	Primarily zoophagic, and will opportunistically feed on humans. ^{29,35,83}
Feeding location	Unknown.
Resting habits	Unknown.
Flight range	Unknown.
Vector status	<i>Plasmodium</i> spp. (secondary vector). ²⁵

Anopheles oreios Bangs & Harbach 2015

Distribution of *Anopheles oreios* throughout the Pacific Islands



Photo source:
Stephen Doggett

Bionomics of *Anopheles oreios*

Subgenus	<i>Cellia</i>
Distribution	The distribution is patchy and restricted to highland river valleys and plains, at elevations above 1000 m (generally >1500 m), along the central range for high mountains on the main island of New Guinea. ^{67,84-86} Formerly <i>Anopheles farauti</i> 6.
Aquatic habitats	Found in natural and man-made sunlit or partially shaded habitats containing freshwater, such as ground pools, permanent swamps and marshes, flooded meadows, ditches, pits, pig wallows and fish ponds. ⁸⁴
Feeding times	Active host-seeking occurs predominantly in the first half of the evening (nocturnal). ⁸⁴
Blood meal hosts	Indiscriminate and opportunistic blood-feeders on humans and other animals, such as pigs and dogs. ⁸⁴
Feeding location	Predominantly exophagic, infrequently entering houses to feed. ⁸⁴
Resting habits	Predominantly exophilic, infrequently resting in houses. ⁸⁴
Flight range	Nothing about dispersal is known or recorded, but <i>An. oreios</i> females likely remain close to their larval habitats as long as acceptable blood sources are relatively near (50–100 m distance). ⁸⁴
Vector status	<i>Plasmodium</i> spp. (secondary vector, suspected to be an important vector in the highlands above 1500 m). ⁸⁴

Anopheles punctulatus Dönitz 1901

Distribution of *Anopheles punctulatus* throughout the Pacific Islands

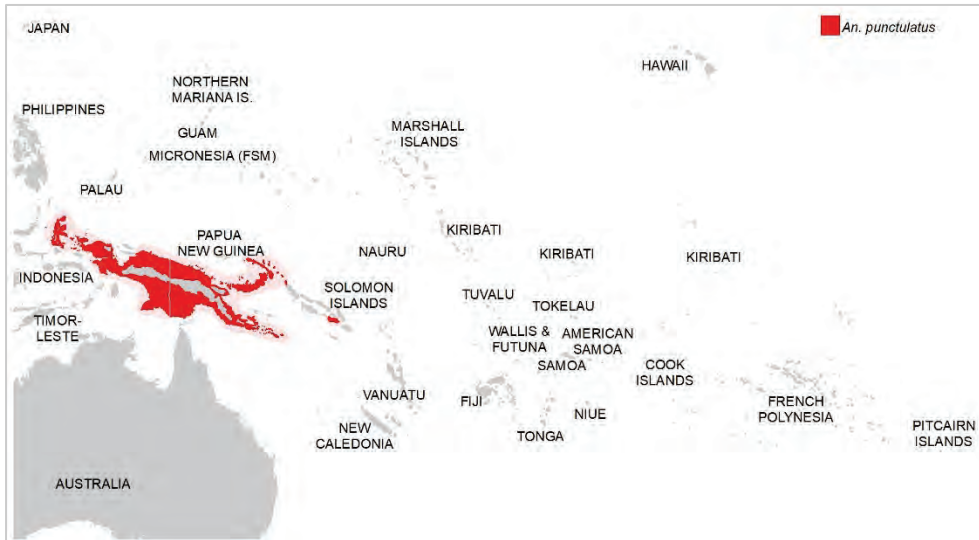


Photo source:
Walter Reed
Biosystematics Unit

Bionomics of *Anopheles punctulatus*

Subgenus	<i>Cellia</i>
Distribution	Found in the Moluccas, New Guinea, New Britain, New Ireland and Buka. In the Solomon Islands, the distribution is restricted to Guadalcanal. Most common in lowland areas. ^{27,28,33,37,50,67,70,74,76,77,87-90}
Aquatic habitats	The aquatic habitats are mostly small transient sites exposed to sunlight with little vegetation, such as pools in rivers or streambeds, wheel ruts, puddles, hoof and footprints, pig wallows and shallow drains. ⁶⁷
Feeding times	Feeds throughout the night, with peak activity between 2200–0200 (nocturnal). ⁵³
Blood meal hosts	Highly anthropophilic. ^{35,55}
Feeding location	Readily feeds both indoors and outdoors; endophagy varies by location.
Resting habits	While they may rest indoors for a short time after feeding indoors, most resting is outdoors.
Flight range	Unknown.
Vector status	<i>Plasmodium</i> spp. (primary vector), ^{15,25,53,79} <i>Wuchereria bancrofti</i> (primary vector). ^{62,65,80} Epidemiological importance in the Solomon Islands is limited due to restricted range.

Aedes aegypti (Linnaeus) 1762

Distribution of *Aedes aegypti* throughout the Pacific Islands

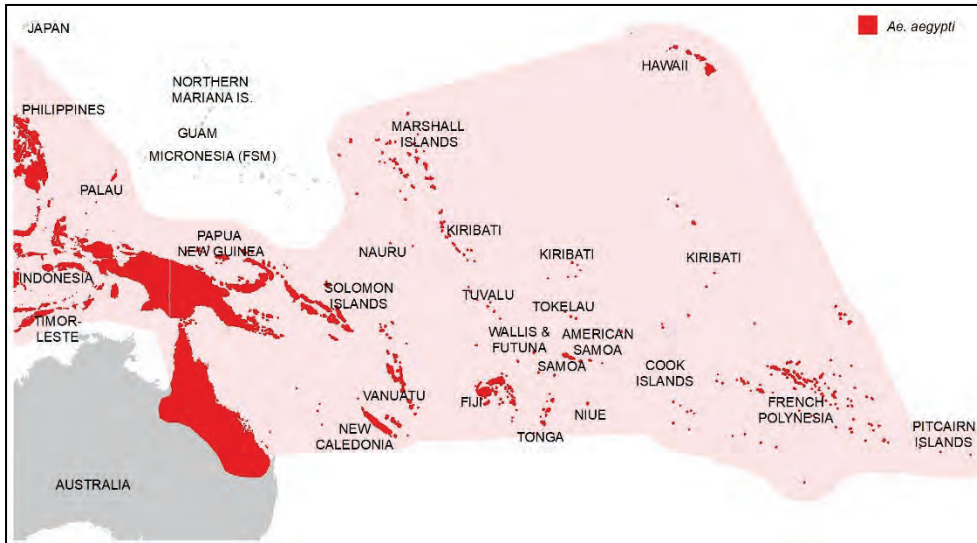


Photo source:
Stephen Doggett

Bionomics of *Aedes aegypti*

Subgenus	<i>Stegomyia</i>
Distribution	A highly invasive species now found across the globe and the Pacific. ^{31,91-107} <i>Aedes aegypti</i> is commonly found in urban and semiurban environments in close association with humans. ^{24,27}
Aquatic habitats	Inhabits artificial containers (e.g. tanks, water storage containers, tyres, pot-plant bases, buckets and other containers around or inside homes). ^{94,97,100,101,106,108} Can also utilise natural sites (e.g. bromeliads). Larvae found only in freshwater.
Feeding times	Feeds during daylight hours, usually more active in the early morning or late afternoon (diurnal and crepuscular). ^{24,103,108} Recently reported to bite after sundown until midnight in the Solomon Islands (Butafa, unpublished data).
Blood meal hosts	Prefers humans, but occasionally bites dogs, cats and other animals. ³⁴
Feeding location	Tends to feed more commonly indoors, as well as in shaded locations near houses. ¹⁰⁸
Resting habits	Rests indoors in dark shady areas below 1.5 m from the floor, and shady areas just outside the house, e.g. under outdoor furniture. ¹⁰⁹
Flight range	Limited flight dispersal – usually only 50–100 m. ¹¹⁰⁻¹¹²
Medical importance	Dengue, chikungunya, Zika viruses (primary vector) as well as yellow fever, Ross River (secondary vector) and numerous other arboviruses. ^{24,108,113-119} Is a competent vector of <i>Dirofilaria immitis</i> (dog heart worm) in the Americas, ¹²⁰ but the strong anthropogenic behaviour of this species may limit its role in transmission. ¹⁰³

Aedes albolineatus (Theobald) 1904

Distribution of *Aedes albolineatus* throughout the Pacific Islands

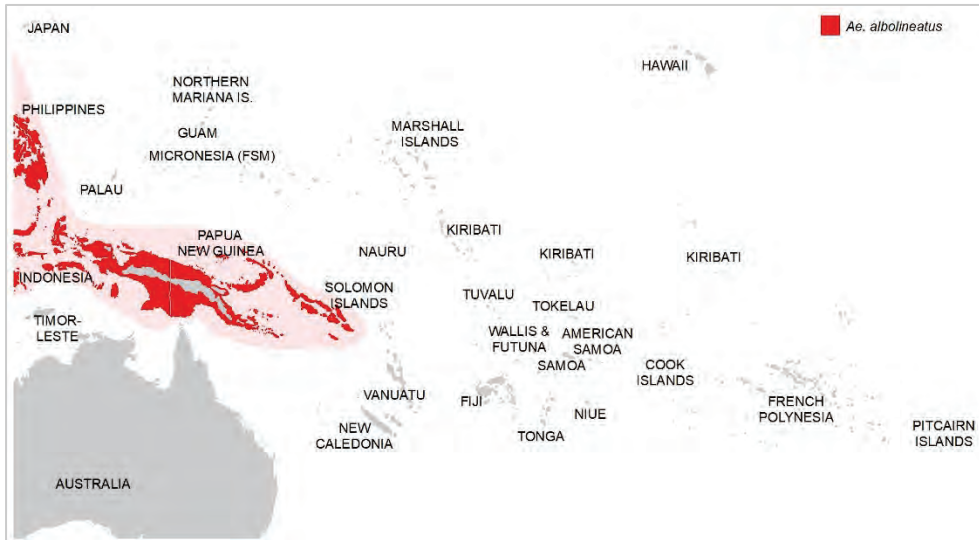


Photo source: Monarch

Bionomics of *Aedes albolineatus*

Subgenus	<i>Scutomyia</i>
Distribution	Found in southeast Asia and in the Pacific in Papua New Guinea and the Solomon Islands (excepting Temotu). ^{27,121,122}
Aquatic habitats	Commonly found in natural containers including tree holes, coconut husks, leaf axils and bamboo stumps. Less commonly found in artificial containers. ¹²¹
Feeding times	Unknown.
Blood meal hosts	Zoophagic and rarely attracted to humans. ¹²¹
Feeding location	Feeds outdoors.
Resting habits	Rests outdoors in vegetation.
Flight range	Unknown.
Medical importance	Unlikely a vector of human pathogens because of its zoophagic biting habits. ¹⁷

Aedes albopictus (Macquart) 1903

Distribution of *Aedes albopictus* throughout the Pacific Islands

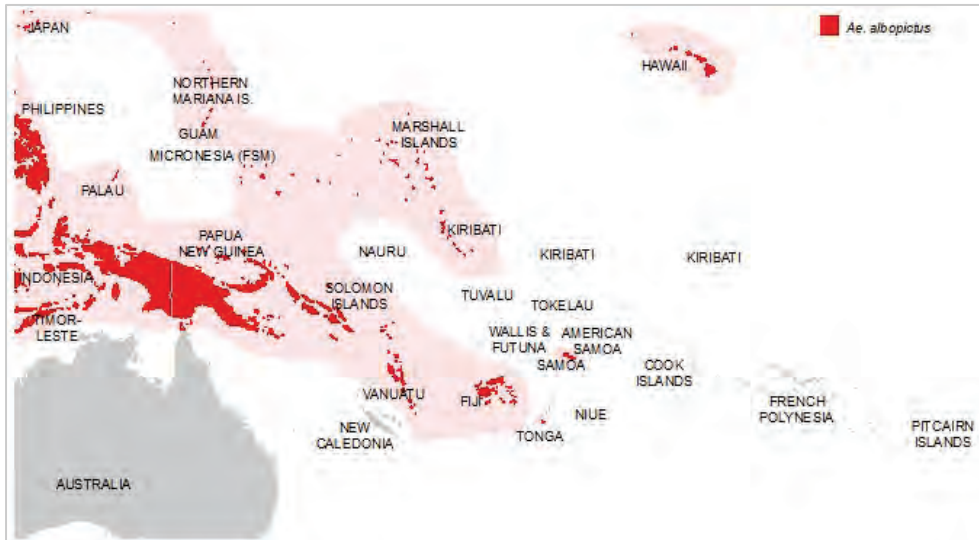


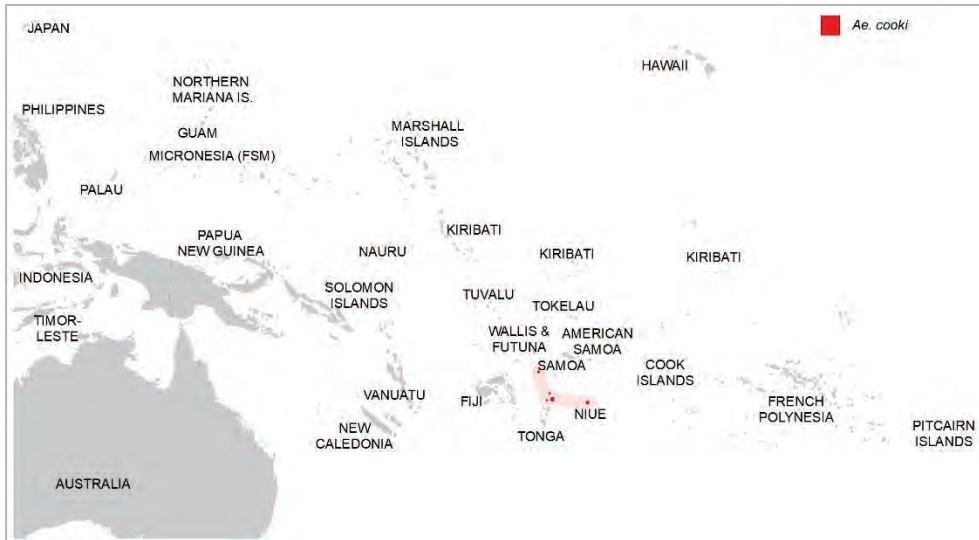
Photo source:
Stephen Doggett

Bionomics of *Aedes albopictus*

Subgenus	<i>Stegomyia</i>
Distribution	A highly invasive species now widely distributed across the globe and in the Pacific. ^{24,27,91,94,97,105,123-126} Often found in vegetated and shaded areas close to human habitation.
Aquatic habitats	Inhabits a broad range of containers, from natural sites (e.g. bamboo stumps, bromeliads, coconuts and tree holes) to artificial containers. ^{94,105,127,128} Has displaced <i>Ae. aegypti</i> in some locations.
Feeding times	Feeds during daylight hours, usually more active in the early morning or late afternoon (crepuscular and diurnal). ¹²⁷ Recently reported to bite after sundown until midnight in the Solomon Islands (Butafa, unpublished data).
Blood meal hosts	Opportunistically feeds on humans and other animals. ¹²⁷ Is often a significant nuisance pest.
Feeding location	Tends to feed more outdoors. ¹²⁷
Resting habits	Rests outdoors on vegetation and also indoors. ¹²⁹
Flight range	Average flight range is 50–200 m. ^{112,130}
Medical importance	Dengue, chikungunya and Zika viruses (secondary vector, can be an important vector in the absence of <i>Ae. aegypti</i>) ¹³¹⁻¹³³ as well as Ross River (secondary vector), ^{116,134} Japanese encephalitis (secondary vector), ¹³⁴ yellow fever and numerous other arboviruses. ^{127,129} A vector of <i>Dirofilaria immitis</i> (dog heart worm). ^{133,135}

Aedes cooki Belkin 1962

Distribution of *Aedes cooki* throughout the Pacific Islands



Bionomics of *Aedes cooki*

	<i>Stegomyia</i>
Distribution	Tonga (Vava'u and Niufo'ou) and Niue. ^{104,121,132,136,137} Often found on the periphery of villages rather in the centre. ¹³⁶
Aquatic habitats	Inhabits a wide range of natural sites including tree holes, leaf axils, coconuts, crab holes and rock holes, as well as artificial containers such as canoes, tyres and drums. ¹³⁶
Feeding times	Feeds during daylight hours (diurnal). ¹³⁶
Blood meal hosts	Commonly feeds on humans. ¹³⁶
Feeding location	Tends to feed more commonly indoors ¹³⁶
Resting habits	Commonly rests indoors. ¹³⁶
Flight range	Unknown
Medical importance	<i>Wuchereria bancrofti</i> (secondary vector, an important vector within its range), ^{132,138} dengue virus (secondary vector, an important vector within its range) ¹²¹ and <i>Dirofilaria immitis</i> (dog heart worm). ¹²⁶

Aedes fijiensis Marks 1947

Distribution of Aedes fijiensis throughout the Pacific Islands

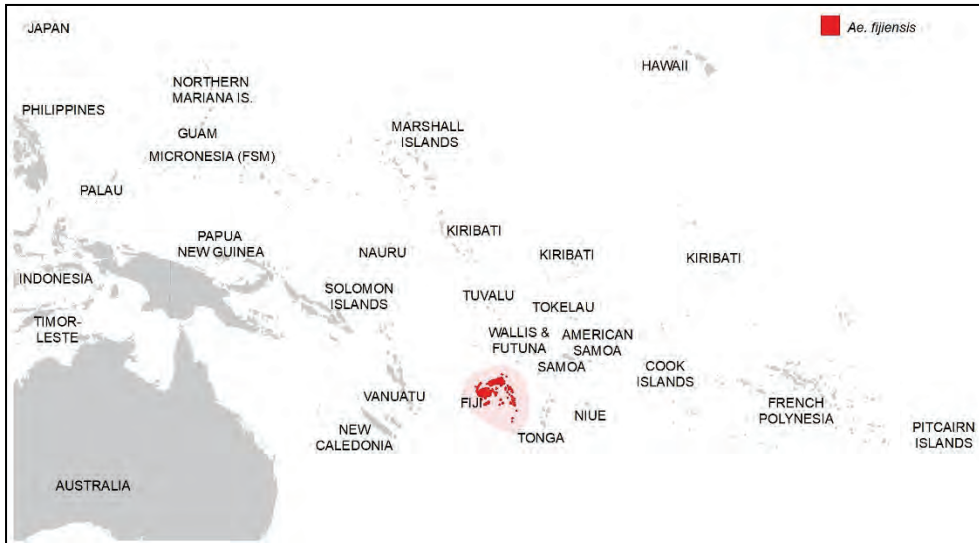


Photo source:
Walter Reed
Biosystematics Unit

Bionomics of Aedes fijiensis

Subgenus	<i>Finlaya</i>
Distribution	Found only in Fiji. ^{92,139-141} Easily confused with other members of the <i>kochi</i> group. Is often a significant nuisance pest. ²⁴ Most commonly found near <i>Pandanus</i> swamps. ¹⁴⁰
Aquatic habitats	The aquatic habitats are the leaf axils of plants, especially <i>Pandanus</i> sp. (screw palm) and <i>Colocasia</i> (taro). ^{139,142}
Feeding times	Feeds during the night (crepuscular and nocturnal). ²⁴
Blood meal hosts	Feeds on humans.
Feeding location	Readily bites indoors or outdoors.
Resting habits	Likely rests mainly outdoors.
Flight range	Unknown.
Vector status	<i>Wuchereria bancrofti</i> (secondary vector due to restricted distribution). ¹⁴¹⁻¹⁴³

Aedes guamensis Farner & Bohart 1944

Distribution of Aedes guamensis throughout the Pacific Islands

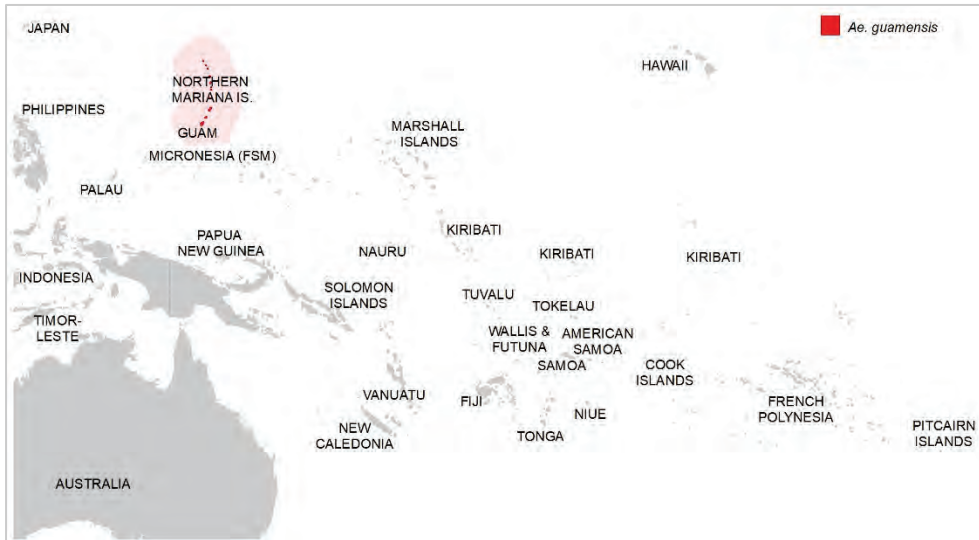


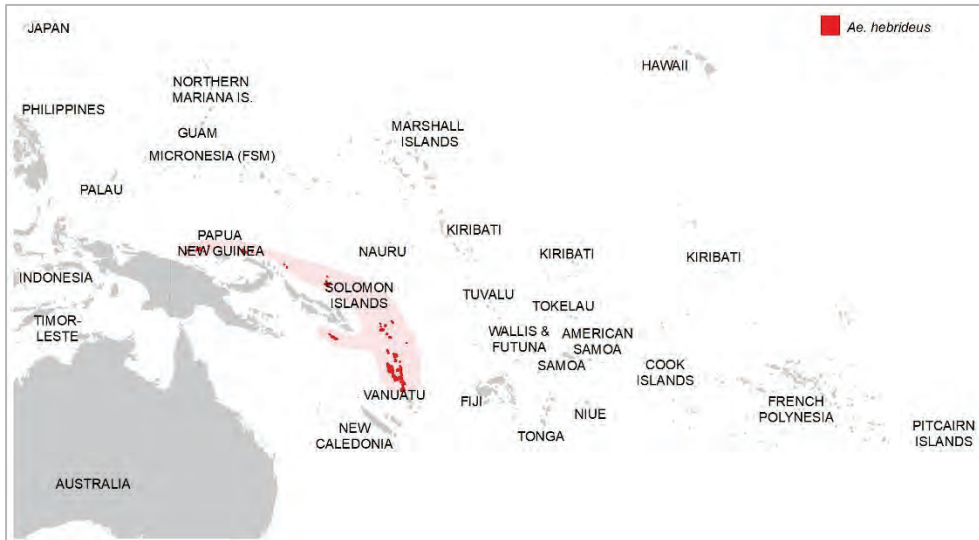
Photo source: Monarch

Bionomics of Aedes guamensis

Subgenus	<i>Stegomyia</i>
Distribution	Guam and the Northern Mariana Islands, often abundant in forested areas. ^{18,126}
Aquatic habitats	A range of natural sites including tree holes and coconuts, as well as artificial containers such as water drums and tanks. ^{126,144}
Feeding times	Observations on feeding time are limited. Has been observed feeding in the evening (crepuscular). ¹⁴⁵
Blood meal hosts	Primarily feeds on animals. Has been recorded to feed on humans. ¹⁴⁶
Feeding location	Outdoors.
Resting habits	Outdoors in vegetation.
Flight range	Unknown.
Medical importance	<i>Dirofilaria immitis</i> (dog heart worm). ¹⁴⁷ Unlikely a vector of human pathogens because of its zoophagic biting habitats.

Aedes hebrideus Edwards 1962

Distribution of Aedes hebrideus throughout the Pacific Islands

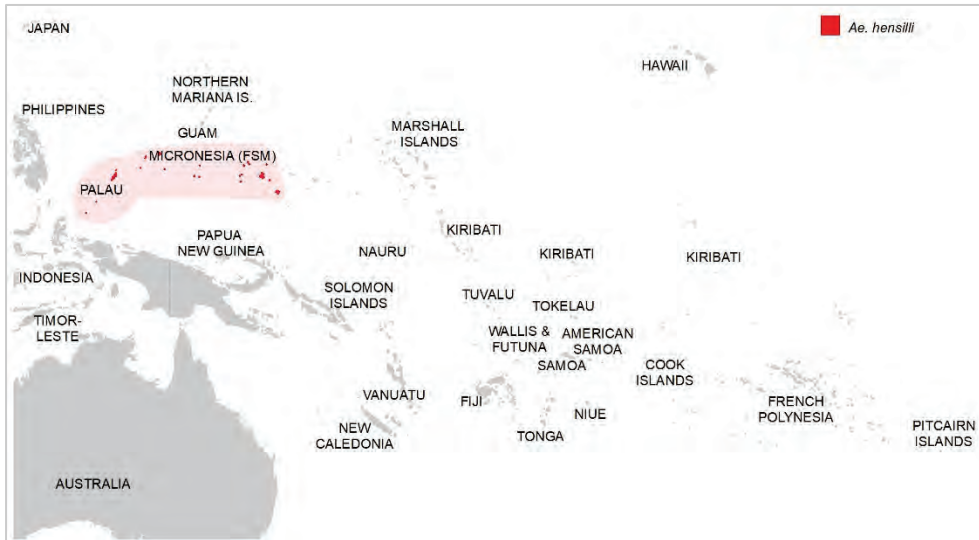


Bionomics of Aedes hebrideus

Subgenus	<i>Stegomyia</i>
Distribution	The outer lying islands north of Papua New Guinea (Wuvulu, Nuguria), Solomon Islands (Rennell, Bellona, Ontong Java, Sikaiana, Temotu) and Vanuatu (excepting Tafea). ^{104,121,132,144,148-151} Considered a semi-domestic species. ^{104,121,152} Very similar to <i>Aedes scutellaris</i> .
Aquatic habitats	Commonly found in natural containers including tree holes, coconut husks and shells, as well as a wide variety of artificial containers. ¹⁴⁴
Feeding times	Feeds during daylight hours (diurnal). ¹²¹
Blood meal hosts	Commonly feeds on humans. ¹²¹
Feeding location	Indoors and outdoors. ¹⁵³
Resting habits	Outdoors in vegetation.
Flight range	Unknown.
Medical importance	Dengue virus (secondary vector). ^{121,132} Inhospitable to <i>Wuchereria bancrofti</i> . ^{65,154}

Aedes hensilli Farner 1945

Distribution of *Aedes hensilli* throughout the Pacific Islands

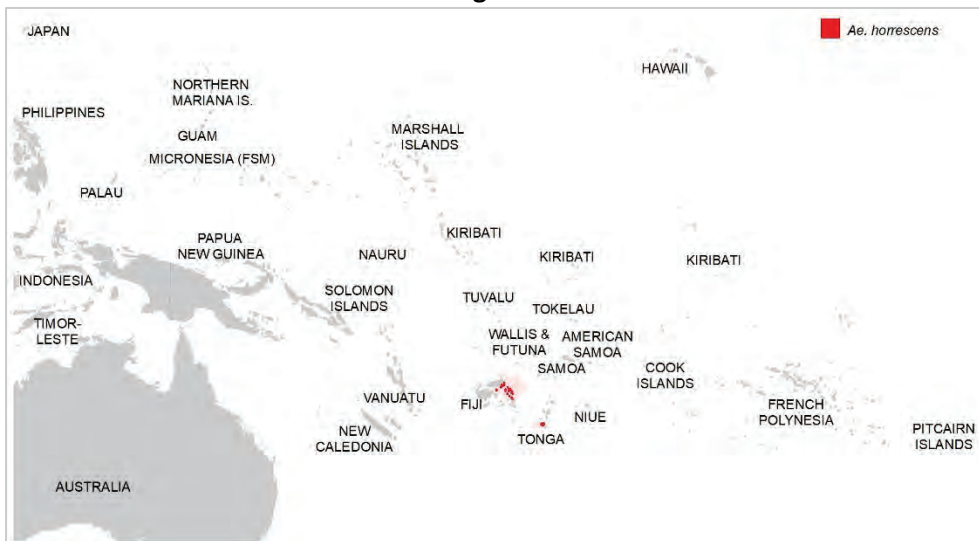


Bionomics of *Aedes hensilli*

Subgenus	<i>Stegomyia</i>
Distribution	Palau and parts of the Federated States of Micronesia (Yap and Chuuk). Can be abundant within its range. ^{132,155}
Aquatic habitats	Artificial containers (e.g. cans and buckets) and natural sites (e.g. coconut shells). ¹⁵⁶⁻¹⁵⁸
Feeding times	Daylight hours, usually more active in the early morning or late afternoon (crepuscular). ¹⁵⁶
Blood meal hosts	Feeds on humans and monkeys. ¹⁵⁹
Feeding location	Feeds either indoors or outdoors.
Resting habits	Generally rests outdoors on vegetation.
Flight range	Unknown.
Medical importance	Dengue, chikungunya and Zika viruses (secondary vector, an important vector within its range). ^{19,156}

Aedes horrescens Edwards 1935

Distribution of *Aedes horrescens* throughout the Pacific Islands



Bionomics of *Aedes horrescens*

Subgenus	<i>Stegomyia</i>
Distribution	Fiji (Taveuni, Makogai, Vanua Balavu) ^{121,132,160} and Tonga (Tongatapu). ^{101,105} Found in forested areas.
Aquatic habitats	Primarily inhabits tree holes but will also utilise coconut shells and occasionally artificial containers. ^{105,121,161}
Feeding times	Unknown.
Blood meal hosts	Primarily feeds on animals (zoophagic). Limited feeding on humans has been observed. ¹⁴⁰
Feeding location	Outdoors.
Resting habits	Outdoors in vegetation.
Flight range	Unknown.
Medical importance	Has limited opportunity to transmit pathogens to humans because of its zoophagic biting habitats. Inhospitable to <i>Wuchereria bancrofti</i> . ^{142,162}

Aedes kesseli Huang & Hitchcock 1980

Distribution of *Aedes kesseli* throughout the Pacific Islands



Bionomics of *Aedes kesseli*

Subgenus	<i>Stegomyia</i>
Distribution	Tonga (Niuas). ^{132,136,137}
Aquatic habitats	Inhabits an extremely wide range of aquatic habitats, including natural containers (e.g. tree holes, coconut shells, rock holes, crab holes) as well as ground pools, swamps and marshes, and also artificial containers (e.g. cans, buckets and water tanks). ¹³⁶
Feeding times	Feeds during daylight hours (diurnal). ¹³⁶
Blood meal hosts	Commonly feeds on humans. ¹³⁶
Feeding location	Feed both indoors and outdoors with the highest densities outdoors at the periphery of villages. ¹³⁶
Resting habits	Generally, rests outdoors on vegetation.
Flight range	Unknown.
Medical importance	<i>Wuchereria bancrofti</i> (secondary vector, an important vector within its range), dengue (secondary vector, an important vector within its range) as well as <i>Dirofilaria immitis</i> (dog heart worm). ^{132,136}

Aedes kochi (Dönitz) 1901

Distribution of *Aedes kochi* throughout the Pacific Islands

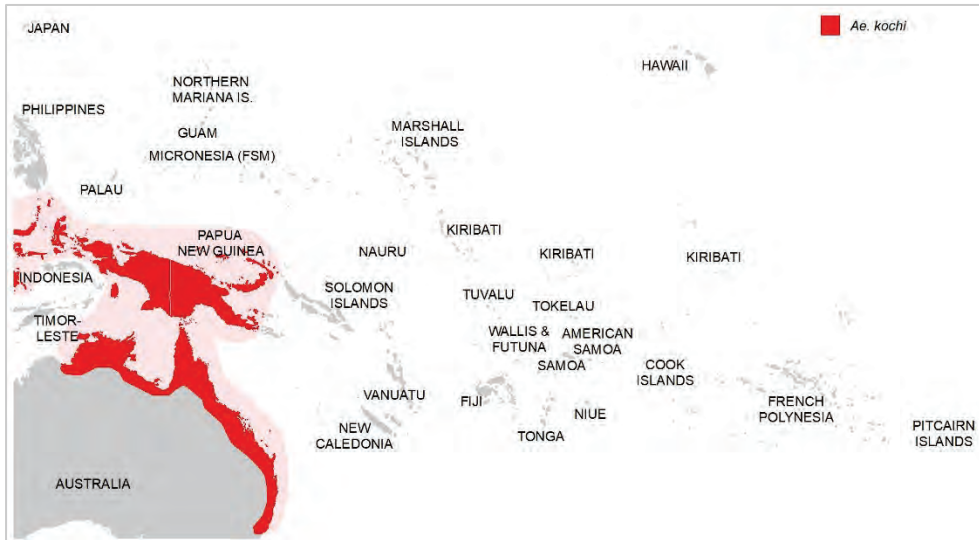


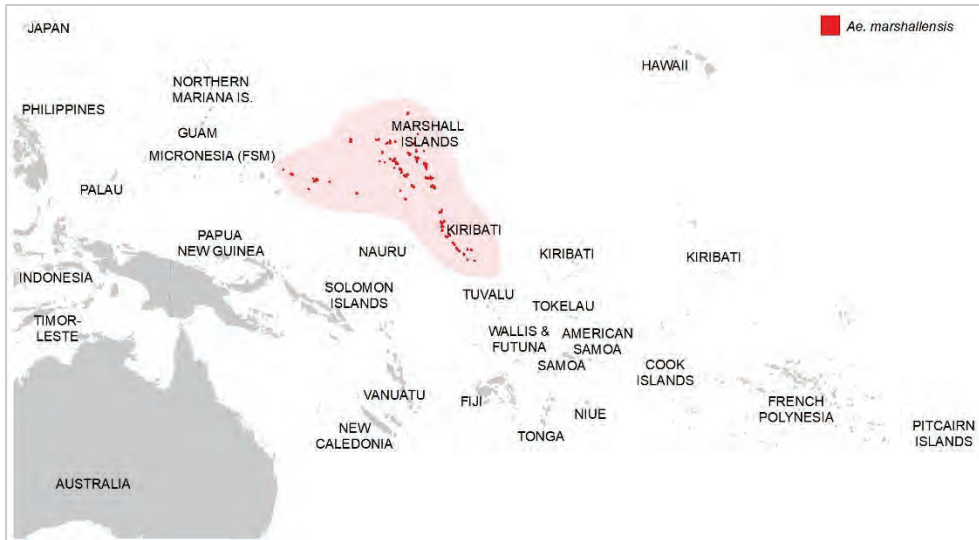
Photo source:
Stephen Doggett

Bionomics of *Aedes kochi*

Subgenus	<i>Finlaya</i>
Distribution	Found in the Moluccas, New Guinea and Australia. ^{27,81,139,141} Easily confused with other members of the kochi group.
Aquatic habitats	The aquatic habitats include leaf axils of a wide variety of plants, including <i>Alocasia</i> , banana, <i>Crinum</i> , pineapple, <i>Pandanus</i> , sago palm, and taro. ²⁴
Feeding times	Peak feeding times vary by location. In New Guinea they readily bite day and night, whereas in New Britain they only bite at night. In Australia, biting starts at 1500, and peaks from dusk until 2300. ^{24,139}
Blood meal hosts	Feeds on humans and has been reported to feed on horses. ^{24,139}
Feeding location	Readily bites indoors or outdoors.
Resting habits	Likely rests mainly outdoors.
Flight range	Unknown.
Vector status	Ross River virus and <i>Wuchereria bancrofti</i> (secondary vector). ^{24,66,139,141,163} Epidemiological importance of this species is limited due to its specific aquatic habitats that restrict the distribution of this species.

Aedes marshallensis (Stone & Bohart) 1944

Distribution of *Aedes marshallensis* throughout the Pacific Islands



Bionomics of *Aedes marshallensis*

Subgenus	<i>Stegomyia</i>
Distribution	Throughout the Marshall Islands and parts of the Federated States of Micronesia (Pohnpei and Kosrae) and Kiribati (Gilbert Islands). ^{96,121,132,145,161} Common in heavily vegetated areas. ¹⁶⁴
Aquatic habitats	Commonly found in natural containers including tree holes, coconut husks and shells, as well as a wide variety of artificial containers including barrels and wells. ^{121,165}
Feeding times	Feeds during daylight hours (diurnal). ^{121,166}
Blood meal hosts	Commonly feeds on humans. ¹⁶¹
Feeding location	Feeds outdoors. ¹⁴⁵
Resting habits	Rests outside in cool damp places. ¹⁴⁵
Flight range	Unknown.
Medical importance	<i>Wuchereria bancrofti</i> (secondary vector), dengue (secondary vector). ^{165,167}

Aedes notoscriptus (Skuse) 1889

Distribution of *Aedes notoscriptus* throughout the Pacific Islands

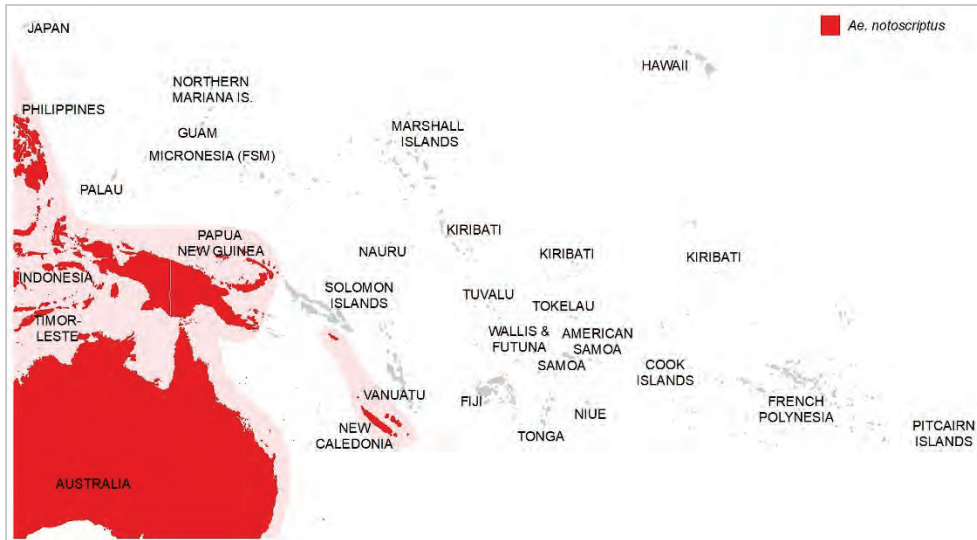


Photo source:
Stephen Doggett

Bionomics of *Aedes notoscriptus*

Subgenus	<i>Rampamyia</i>
Distribution	Native to Australia, and now established in New Zealand, Indonesia, Papua New Guinea, the Solomon Islands (Rennell and Bellona), New Caledonia and California, United States. ^{24,27,31,121,125,150,151,168} Is often a significant nuisance pest.
Aquatic habitats	Aquatic habitats include a variety of natural containers (leaf axils, rock pools, tree holes, bamboo stumps and coconut husks) and a variety of large and small artificial containers made of cement, wood and metal. ¹⁶⁹
Feeding times	Typically blood-feeds in the early morning or late afternoon (crepuscular), but can bite at any time during the day or night. ^{31,92,169}
Blood meal hosts	A range of hosts including humans, dogs, brushtail possums, birds, cats, and flying foxes. ^{24,34}
Feeding location	Readily feeds indoors and outdoors in shaded areas. Will seek hosts at ground level and up to 10 m in the canopy. ^{169,170}
Resting habits	Rests outdoors on vegetation. ¹⁶⁹
Flight range	Up to 200 m. ^{31,112}
Vector status	Numerous arboviruses including Ross River virus (secondary vector), ¹⁷¹ as well as <i>Dirofilaria immitis</i> (dog heart worm). ¹⁷² Not a competent vector of dengue, chikungunya or <i>Wuchereria bancrofti</i> . ^{141,173}

Aedes oceanicus Belkin 1962

Distribution of Aedes oceanicus throughout the Pacific Islands

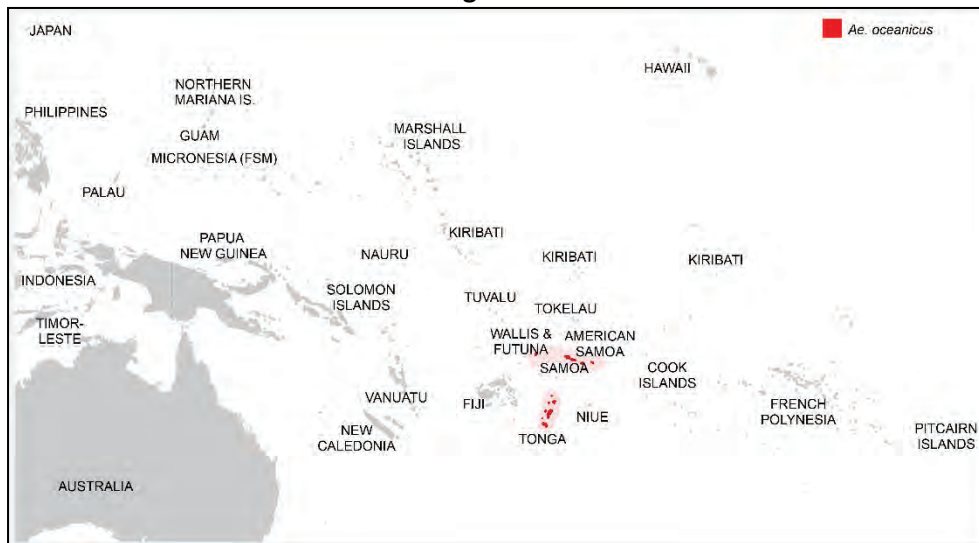


Photo source:
Mark Schmaedick

Bionomics of Aedes oceanicus

Subgenus	<i>Finlaya</i>
Distribution	Found in Tonga ('Eua, Tongatapu, Ha'apai, Vava'u), Samoa (Savai'i, Upolu), American Samoa (Tutuila, Manu'a, Aunu'u) and Wallis. Can be abundant within its range. ^{93,121,138,141} The species was first described in 1962, and in studies prior to that date it was grouped under the name <i>Aedes kochi</i> and later under <i>Aedes samoanus</i> . Currently, all three species (<i>Ae. kochi</i> , <i>Ae. samoanus</i> and <i>Ae. oceanicus</i>) are considered valid, distinct species.
Aquatic habitats	Traditionally found only in plant leaf axils (taro, giant taro, pandanus, pineapple and sago palm), ¹⁷⁴ recently larvae were found in a variety of domestic containers (buckets, plastic and expanded polystyrene containers, cans and tyres). ¹⁷⁵
Feeding times	Blood-feeding begins at dusk and continues throughout the night (crepuscular and nocturnal). ¹³⁸
Blood meal hosts	Will feed on humans and animals.
Feeding location	Bites predominantly outdoors, but will enter houses to feed. ¹³⁸
Resting habits	Rests outdoors. ¹³⁸
Flight range	Unknown.
Vector status	<i>Wuchereria bancrofti</i> (secondary vector in Tonga, inefficient vector in the Samoan Islands). ^{21,141,176-179}

Aedes polynesiensis Marks 1951

Distribution of Aedes polynesiensis throughout the Pacific Islands

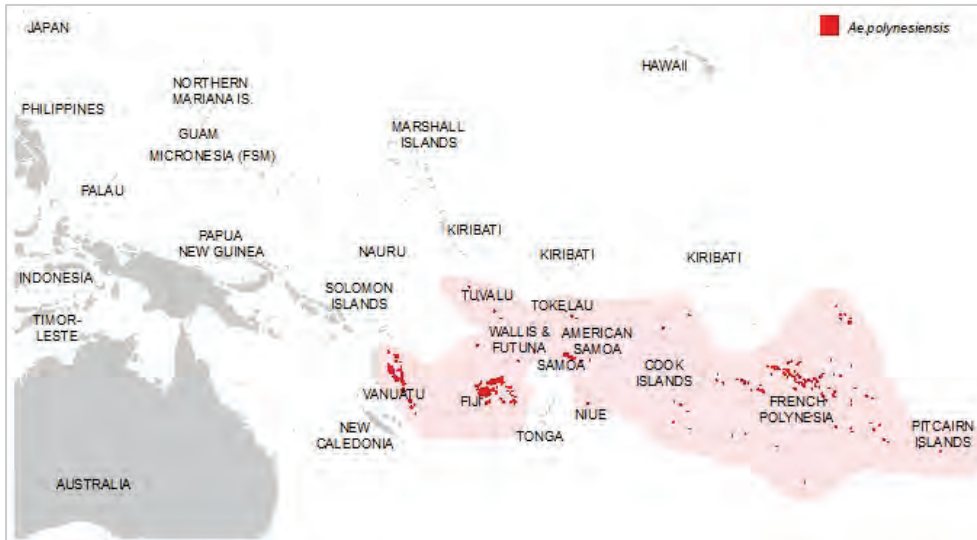


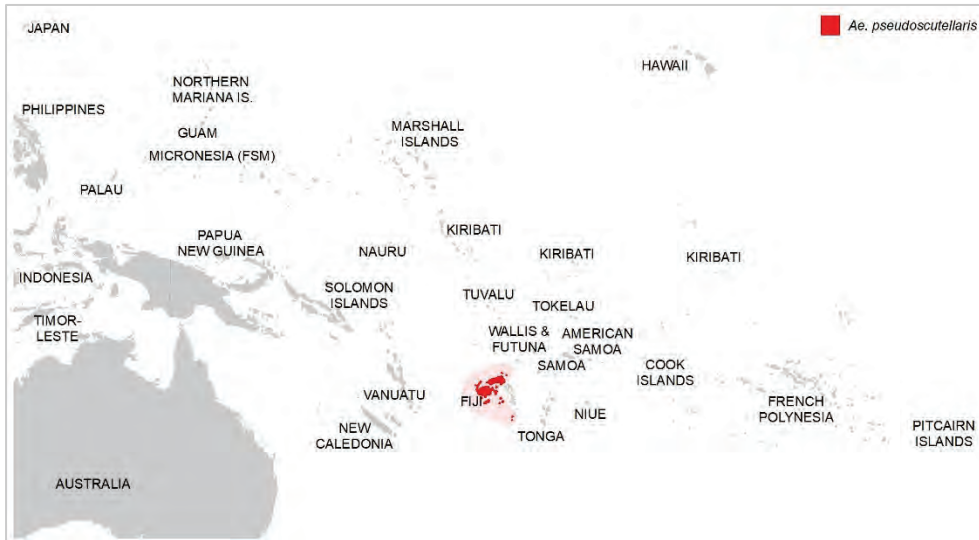
Photo source:
Cook Islands
Biodiversity and
Natural Heritage

Bionomics of Aedes polynesiensis

Subgenus	<i>Stegomyia</i>
Distribution	First described from Taveuni, Fiji, and is common throughout most of Polynesia. ^{39,92,99,121,132,138,140} Recently expanded its range to include Vanuatu. Was called <i>Aedes pseudoscutellaris</i> prior to 1951. Mostly found in coastal areas.
Aquatic habitats	Natural (e.g. tree holes, crab burrows, and coconuts) and artificial (e.g. tyres, pot-plant bases and buckets) containers. ^{100,180} The key aquatic habitats depend on the ecosystem, with natural containers important in bush areas, and artificial containers important in villages.
Feeding times	Feeds during daylight hours, usually more active in the early morning or late afternoon (crepuscular and diurnal); a low level of feeding may continue throughout the night. ^{92,138,181}
Blood meal hosts	Opportunistically feeds on humans and other animals. ¹³⁸
Feeding location	Typically feeds outdoors. ¹⁰³
Resting habits	Generally rests outdoors on vegetation. ¹⁸¹
Flight range	Limited flight dispersal, < 100 m. ^{182,183}
Medical importance	<i>Wuchereria bancrofti</i> (primary vector), ^{21,92,103,132,181,184,185} dengue, chikungunya, Zika (secondary vector) ^{117,132} and Ross River viruses (secondary vector) ^{39,119,186} as well as <i>Dirofilaria immitis</i> (dog heart worm). ^{103,181,185,187}

Aedes pseudoscutellaris (Theobald) 1901

Distribution of *Aedes pseudoscutellaris* throughout the Pacific Islands

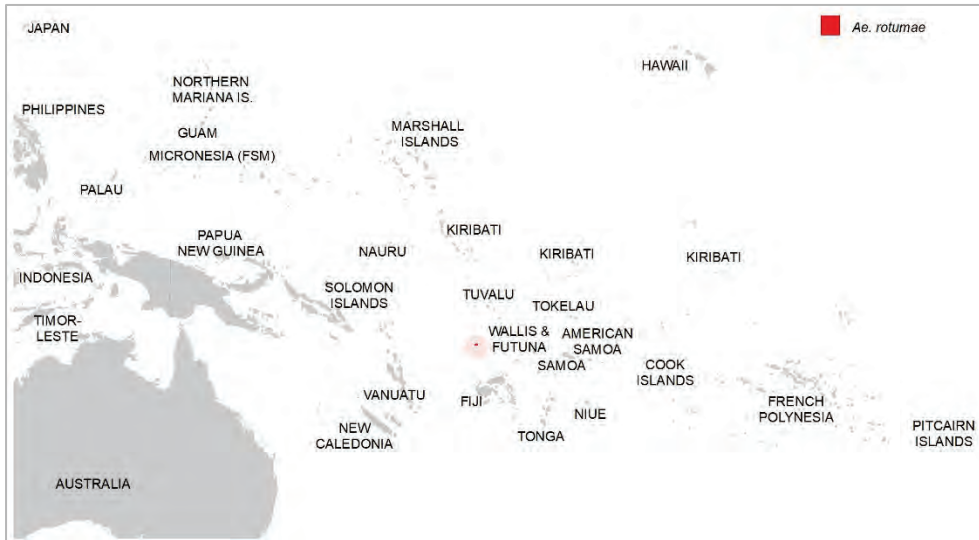


Bionomics of *Aedes pseudoscutellaris*

Subgenus	<i>Stegomyia</i>
Distribution	Only in Fiji (Vanua Levu, Viti Levu, Kandavu, Lau group). ^{92,104,121,132,140,148,188,189} Mostly found inland (>500 m from the coast).
Aquatic habitats	Commonly found in natural containers including tree holes, coconut husks and shells, as well as a wide variety of artificial containers including barrels and wells. ^{94,121}
Feeding times	Feeds primarily during the daytime (diurnal), although a low level of feeding can continue throughout the night. ⁹²
Blood meal hosts	Commonly feeds on humans. ⁹²
Feeding location	Typically feeds outdoors.
Resting habits	Outdoors on vegetation.
Flight range	Unknown.
Medical importance	<i>Wuchereria bancrofti</i> (primary vector), dengue virus (secondary vector). ^{190,191}

Aedes rotumae Belkin 1962

Distribution of *Aedes rotumae* throughout the Pacific Islands



Bionomics of *Aedes rotumae*

Subgenus	<i>Stegomyia</i>
Distribution	Only on Rotuma Island in Fiji. ^{121,132}
Aquatic habitats	Tree holes, in coconut shells and in tin cans. ¹²¹
Feeding times	Feeds during the day.
Blood meal hosts	Feeds on humans. ¹²¹
Feeding location	Typically feeds outdoors. ¹²¹
Resting habits	Generally rests outdoors on vegetation. ¹²¹
Flight range	Unknown.
Medical importance	<i>Wuchereria bancrofti</i> (secondary vector, an important vector within its range) and dengue virus (secondary vector, an important vector within its range). ^{20,121,132}

Aedes samoanus (Grünberg) 1913

Distribution of *Aedes samoanus* throughout the Pacific Islands

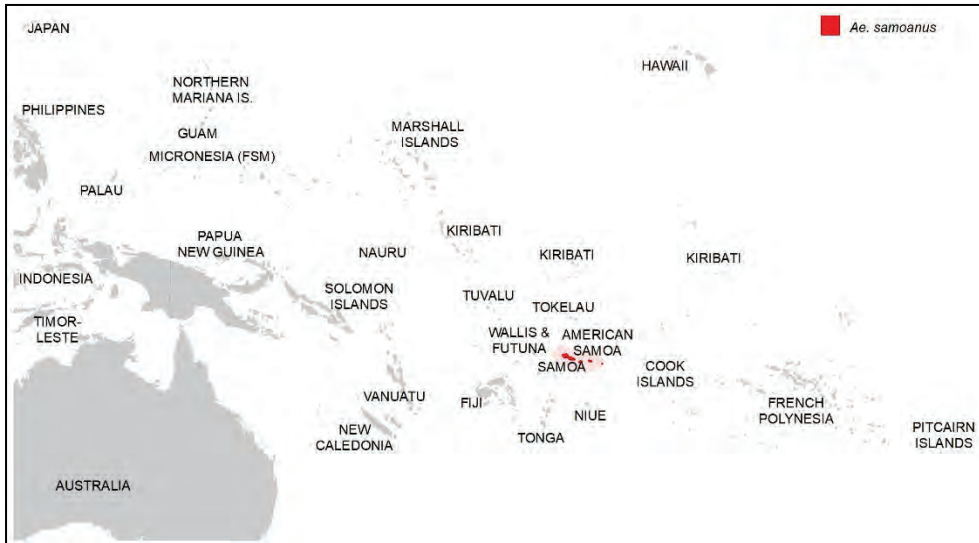


Photo source:
Mark Schmaedick

Bionomics of *Aedes samoanus*

Subgenus	<i>Finlaya</i>
Distribution	Found only in Samoa and American Samoa. ^{138,141} Abundant in villages surrounded by forest, as well as in the forests and plantations. ^{21,192}
Aquatic habitats	Leaf axils of <i>Freycinetia</i> sp. (throughout its range) and pandanus (Samoa only). ^{21,174,192}
Feeding times	Feeds at night time (nocturnal), becoming active after sunset and peaking of activity at 11 pm. ^{21,193}
Blood meal hosts	Commonly feeds on humans. ¹²¹
Feeding location	Tends to feed more outdoors. ²¹
Resting habits	Rests outdoors in vegetation. ²¹
Flight range	Unknown.
Vector status	<i>Wuchereria bancrofti</i> (primary vector). ^{21,141,192,194}

Aedes scutellaris (Walker) 1858

Distribution of Aedes scutellaris throughout the Pacific Islands

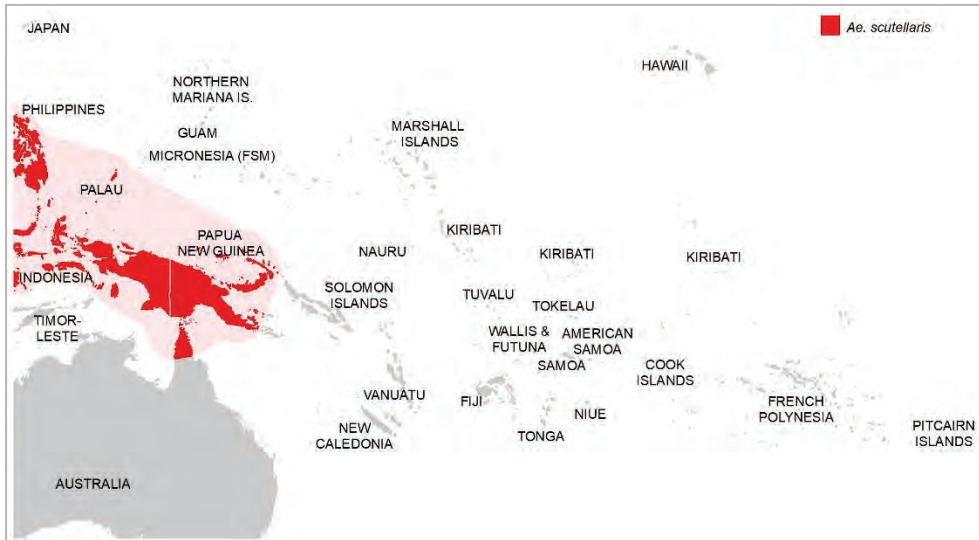


Photo source:
Walter Reed
Biosystematics Unit

Bionomics of Aedes scutellaris

Subgenus	<i>Stegomyia</i>
Distribution	Philippines, Palau, the Moluccas, New Guinea and northern Queensland. ^{121,141,155,195} This species does not naturally occur in the South Pacific; the form reported as <i>scutellaris</i> from this area is <i>hebrideus</i> . ¹²¹ It was detected in New Caledonia in 2016–17, but not since. ¹⁹⁶
Aquatic habitats	Aquatic habitats include coconut shells, tree holes, palm fronds, <i>Pandanus</i> leaf axils, bamboo, and artificial containers such as empty cans. Occasionally, larvae are seen in small ground pools and wells. ^{24,132,197}
Feeding times	The peak biting time is in the early evening (crepuscular), but will bite opportunistically during the day in shaded areas. ¹⁹⁷
Blood meal hosts	Feeds on humans. ¹⁶¹
Feeding location	Indoors or outdoors. ¹⁹⁷
Resting habits	Generally outdoors on vegetation.
Flight range	Unknown.
Vector status	Dengue virus (secondary vector). ^{132,161,198} Inhospitable to <i>Wuchereria bancrofti</i> . ⁶⁶

Aedes tabu Ramalingam & Belkin 1965

Distribution of *Aedes tabu* throughout the Pacific Islands



Bionomics of *Aedes tabu*

Subgenus	<i>Stegomyia</i>
Distribution	Tonga (Tongatapu and Eua). ^{132,136} Is a member of the <i>Ae. tongae s.l.</i> complex. ¹³⁶
Aquatic habitats	Natural containers including tree holes, coconut holes and leaf axils, as well as artificial containers. ^{136,138}
Feeding times	During the day (diurnal). ¹³⁶
Blood meal hosts	Feeds on humans. ¹³⁶
Feeding location	Feed both indoors and outdoors, with the highest densities outdoors at the periphery of villages. ¹³⁶
Resting habits	Generally rests outdoors on vegetation.
Flight range	Unknown.
Vector status	<i>Wuchereria bancrofti</i> (primary vector within its range), ^{21,177} dengue virus (secondary vector, an important vector within its range) ^{136,199} as well as <i>Dirofilaria immitis</i> (dog heart worm). ¹³⁶

Aedes tongae s.s. Edwards 1926

Distribution of *Aedes tongae* s.s. throughout the Pacific Islands



Bionomics of *Aedes tongae* s.s.

Subgenus	<i>Stegomyia</i>
Distribution	Tonga (Ha'apai). ¹³⁶ Abundant in plantations and shady areas of villages. Is a member of the <i>Ae. tongae</i> s.l. complex. ¹³⁶
Aquatic habitats	Natural containers including tree holes, coconut holes and leaf axils, as well as artificial containers. ^{136,138}
Feeding times	During the day in shaded areas (diurnal), with peak biting between 10 am and 12 pm. ^{21,136}
Blood meal hosts	Feeds on humans. ²¹
Feeding location	Indoors or outdoors. ¹³⁶
Resting habits	Outdoors in vegetation. ²¹
Flight range	Unknown.
Vector status	<i>Wuchereria bancrofti</i> (primary vector within its range), ²¹ dengue virus (secondary vector, an important vector within its range) ^{136,199} as well as <i>Dirofilaria immitis</i> (dog heart worm). ¹³⁶

Aedes tutuilae Ramalingam & Belkin 1965

Distribution of *Aedes tutuilae* throughout the Pacific Islands



Bionomics of *Aedes tutuilae*

Subgenus	<i>Finlaya</i>
Distribution	Found only in Samoa and American Samoa. ^{138,141,200}
Aquatic habitats	Leaf axils of pandanus. ^{174,200}
Feeding times	Feeds at night time (nocturnal). ¹³⁸
Blood meal hosts	Does feeds on humans, tendency to feed on animals is unknown. ¹⁹⁴
Feeding location	Does feed outdoors. ¹⁹⁴
Resting habits	Unknown.
Flight range	Unknown.
Vector status	A suspected, but not confirmed, vector of <i>Wuchereria bancrofti</i> . ^{21,179,194}

Aedes upolensis Marks 1957

Distribution of *Aedes upolensis* throughout the Pacific Islands

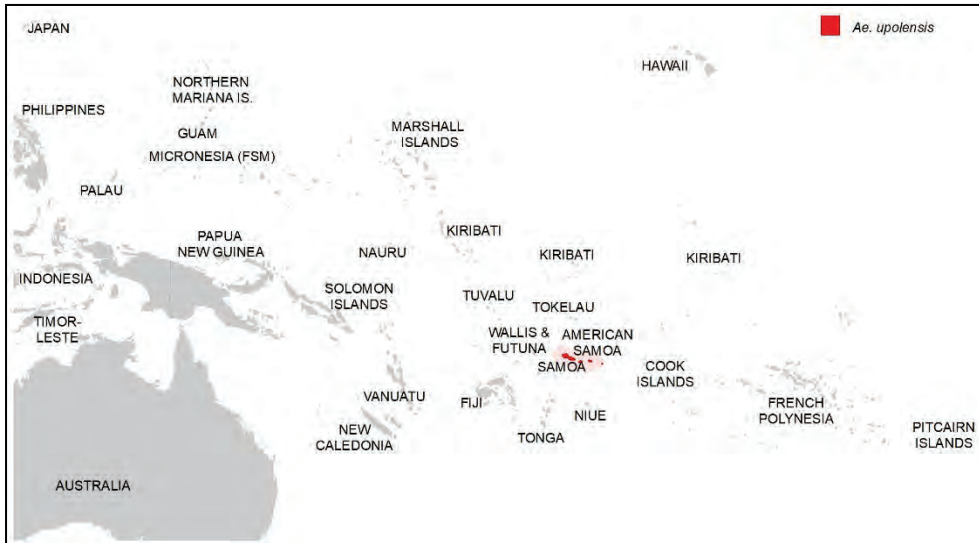


Photo source:
Mark Schmaedick

Bionomics of *Aedes upolensis*

Subgenus	<i>Stegomyia</i>
Distribution	Restricted to the Samoan Islands. Found in dense bush or forested areas, and villages near to the forest. ^{21,132,185,201,202}
Aquatic habitats	Aquatic habitats include tree holes. ^{21,201}
Feeding times	During the day (diurnal). ²¹
Blood meal hosts	Feeds on humans. ^{21,201}
Feeding location	Outdoors.
Resting habits	Outdoors on vegetation.
Flight range	Unknown.
Vector status	<i>Wuchereria bancrofti</i> (secondary vector, an important vector within its range). ^{21,185}

Aedes vexans (Meigen) 1830

Distribution of *Aedes vexans* throughout the Pacific Islands

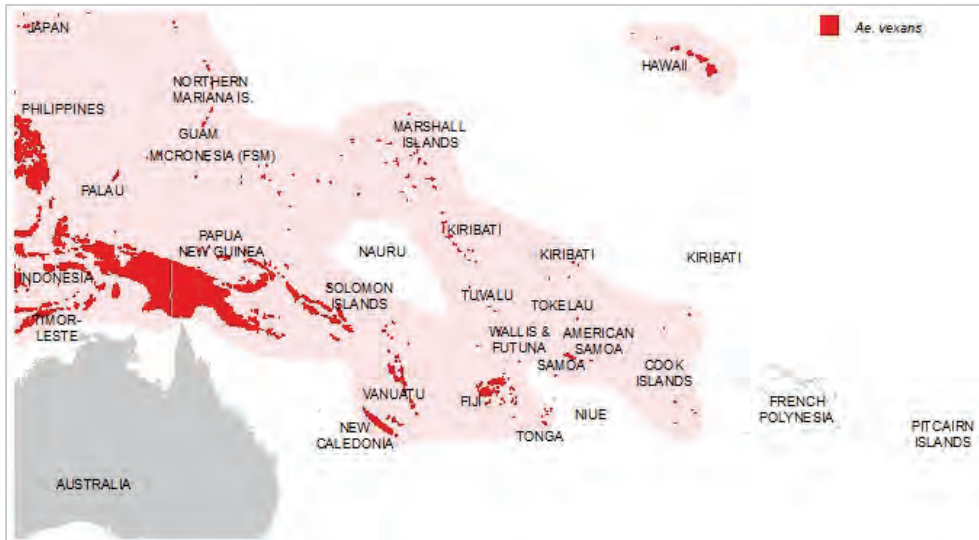


Photo source:
Christine Young

Bionomics of *Aedes vexans*

Subgenus	<i>Aedimorphus</i>
Distribution	One of the most widely distributed species in the world. The subspecies <i>Ae. vexans nocturnus</i> is found in the Australasian and Pacific region. ^{27,92,96,105,121,124,138,140,141,155,203-210} Is often a severe nuisance pest.
Aquatic habitats	Larvae prefer sunlit freshwater habitats, usually temporary in nature such as ground pools, ditches, swamps or marshes, and rarely in containers. Habitats usually have little aquatic vegetation or algae. ^{105,113,211} Larvae can hatch in extremely high numbers.
Feeding times	Females will bite at any time of the day or night, with peak biting starting at sunset and continuing into the night. ⁹²
Blood meal hosts	Opportunistically feeds on a range of mammals including humans. ^{113,211,212}
Feeding location	Will readily feed outdoors or indoors.
Resting habits	Rests outdoors on low vegetation. ²¹²
Flight range	Will disperse up to 14 km from aquatic habitats, with a flight range of up to 1 km/night. ^{112,113}
Vector status	Numerous arboviruses including Japanese encephalitis (competent vector in Asia) ^{213,214} and West Nile. Regarding Zika virus, American populations are susceptible to infection, but dissemination and transmission rates are relatively low. ²¹⁵ <i>Dirofilaria immitis</i> (dog heart worm). ^{204,211} Inhospitable to <i>Wuchereria bancrofti</i> . ^{140,141}

Aedes vigilax (Skuse) 1889

Distribution of *Aedes vigilax* throughout the Pacific Islands

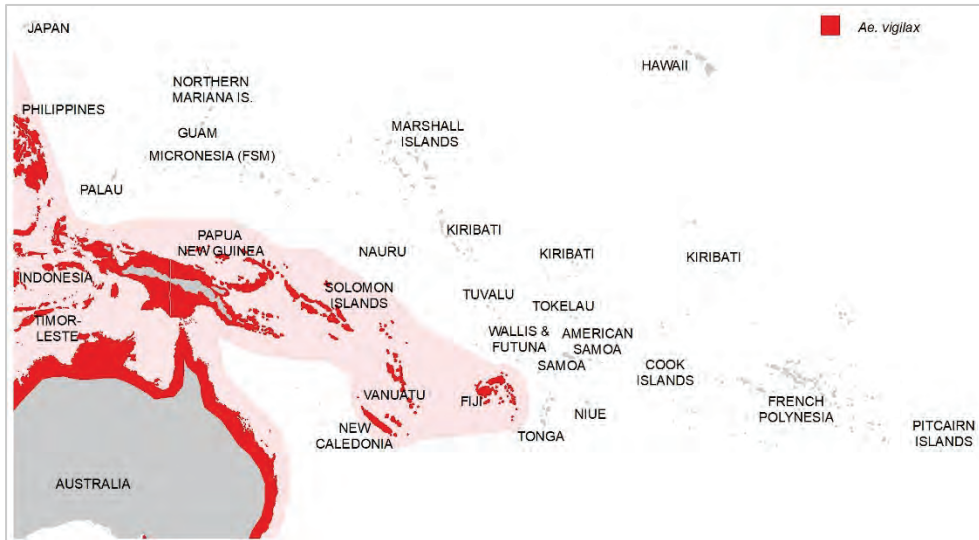


Photo source:
Stephen Doggett

Bionomics of *Aedes vigilax*

Subgenus	<i>Ochlerotatus</i>
Distribution	Widely distributed in salt marshes and mangrove swamps across the Australasian and Pacific region. Found inland where dryland salinity creates suitable habitats. ^{27,31,92,95,121,149,216-218}
Aquatic habitats	Usually tidally influenced saltmarsh and mangrove habitats, but can also use other saline and brackish water habitats, such as flooded sedgeland and coastal forests. ²¹⁹
Feeding times	Females will bite any time of the day or night, with peak biting starting at sunset and continuing into the night. ⁹²
Blood meal hosts	Opportunistically feeds on a range of hosts including humans, dogs, birds, brushtail possums, cats, and flying foxes. ³⁴ Is often a significant nuisance pest.
Feeding location	Will readily feed outdoors or indoors.
Resting habits	Rests outdoors on vegetation.
Flight range	<i>Aedes vigilax</i> is a powerful flier and will travel considerable distances from coastal aquatic habitats to find its hosts (up to 3 km). ²²⁰
Vector status	Numerous arboviruses including Ross River virus (primary vector) ^{221,222} and Japanese encephalitis (secondary vector), ^{6,223} <i>Wuchereria bancrofti</i> (primary vector in New Caledonia, inefficient vector in Vanuatu and Fiji), ^{95,216,217} as well as <i>Dirofilaria immitis</i> (dog heart worm). ²²⁴

Culex annulirostris Skuse 1889

Distribution of Culex annulirostris throughout the Pacific Islands

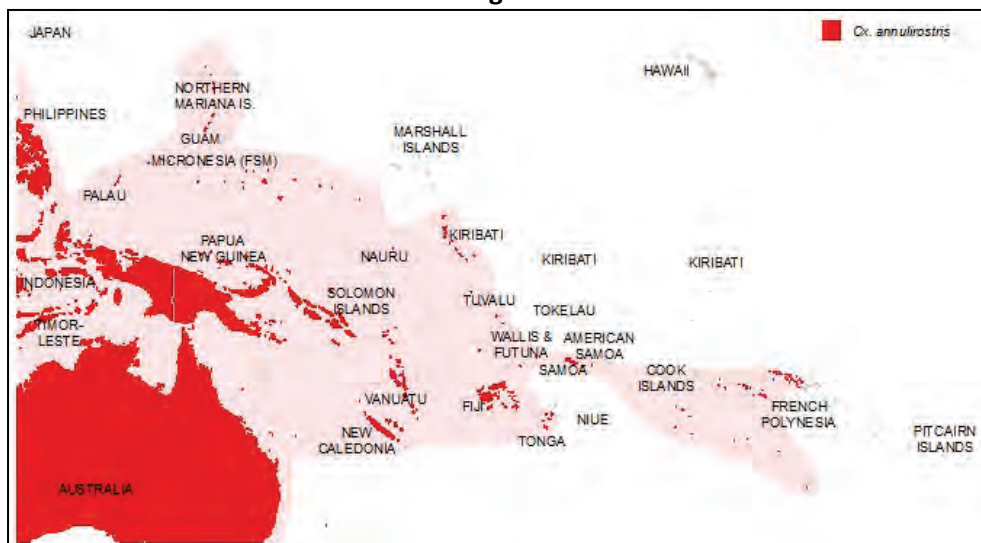


Photo source:
Stephen Doggett

Bionomics of Culex annulirostris

Subgenus	<i>Culex</i>
Distribution	Is found from Indonesia through to the Pacific and as far north as the Philippines. ^{31,39,92,101,138,140,149,225} In the Pacific, <i>Culex annulirostris</i> is found as far east as Kiribati (Gilberts), ²²⁶ Cook Islands (Aitutaki, Rarotonga) and French Polynesia (Society, Austral, Tuamotu). ⁹⁹ Possibly a complex of an unknown number of siblings that may vary in vector competence and vectorial capacity. ³¹ The subspecies in the Marianas Islands is <i>Culex annulirostris marianae</i> . ¹⁴⁵
Aquatic habitats	A wide range of freshwater habitats, ranging from swamps, roadside pools, marshes, temporary and semi-permanent ground waters, natural and artificial containers. ^{31,121}
Feeding times	Mostly at night from dusk to dawn with a peak at midnight, noting that a low level of feeding can occur during the day. ^{31,92,121}
Blood meal hosts	Opportunistically feeds on available animal hosts, including humans, dogs, cats, horses, birds, cats, and fruit bats. ^{24,34} Is often a significant nuisance pest.
Feeding location	Primarily outdoors, but will feed indoors. ¹²¹
Resting habits	Primarily outdoors in vegetation, but has been caught resting indoors. ²²⁷
Flight range	<i>Culex annulirostris</i> has a large flight range (up to 6 km), and the average distance travelled per day, based on mark-recapture experiments, is 2 km. ¹¹²
Medical importance	Numerous arboviruses including Japanese encephalitis (primary vector in Papua New Guinea). ²²⁸ For Ross River virus, has relatively low vector competence in the laboratory, but acts as a primary vector when population densities are high. ^{229,230} <i>Wuchereria bancrofti</i> (important vector in parts of West Papua, Indonesia, inefficient vector throughout the rest of the Pacific), ^{64,163,202,225,231,232} as well as <i>Dirofilaria immitis</i> (dog heart worm). ^{112,187}

Culex bitaeniorhynchus Giles 1901

Distribution of *Culex bitaeniorhynchus* throughout the Pacific Islands

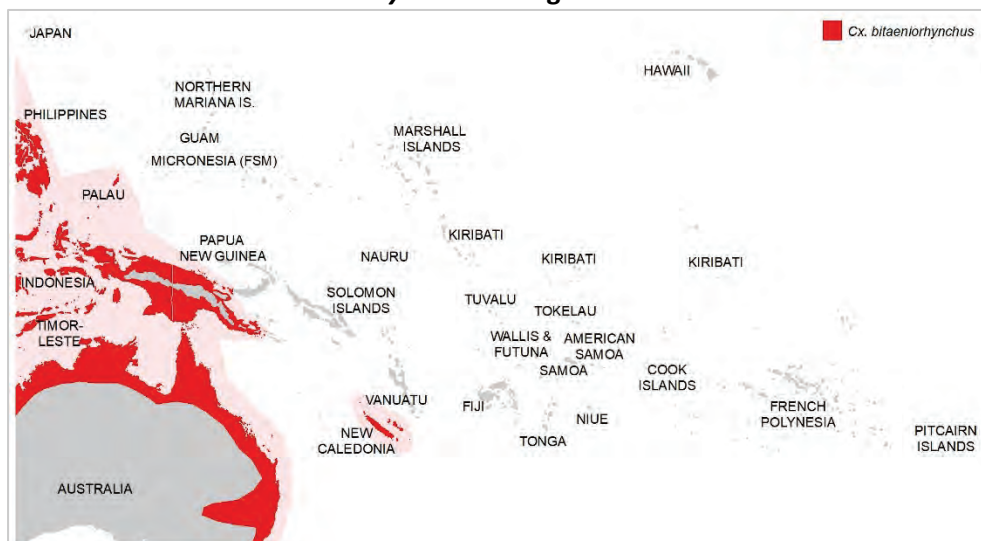


Photo source:
Stephen Doggett

Bionomics of *Culex bitaeniorhynchus*

Subgenus	<i>Oculeomyia</i>
Distribution	A wide global distribution. In the Pacific is naturally found in Australia, New Guinea, and introduced to New Caledonia and Palau. ^{31,225,233}
Aquatic habitats	Permanent freshwater habitats including the margins of wetlands or occasionally in slow-moving streams, as well as wastewater treatment ponds. ^{31,145}
Feeding times	The majority of feeding occurs at dusk and dawn and continues throughout the night (crepuscular and nocturnal). ²⁴
Blood meal hosts	Preferred host is birds, although it will bite humans and a range of other non-avian hosts including cows, pigs and dogs. ²⁴
Feeding location	Outdoors.
Resting habits	Outdoors in vegetation.
Flight range	Unknown.
Medical importance	<i>Wuchereria bancrofti</i> (secondary vector in New Guinea, inefficient vector in New Caledonia). ^{163,225,233,234} Japanese encephalitis (competent vector in Asia, Australian populations can become infected). ^{235,236} Does not play a major role in the transmission of pathogens to humans because of its zoophagic biting habitats.

Culex gelidus Theobald 1901

Distribution of *Culex gelidus* throughout the Pacific Islands

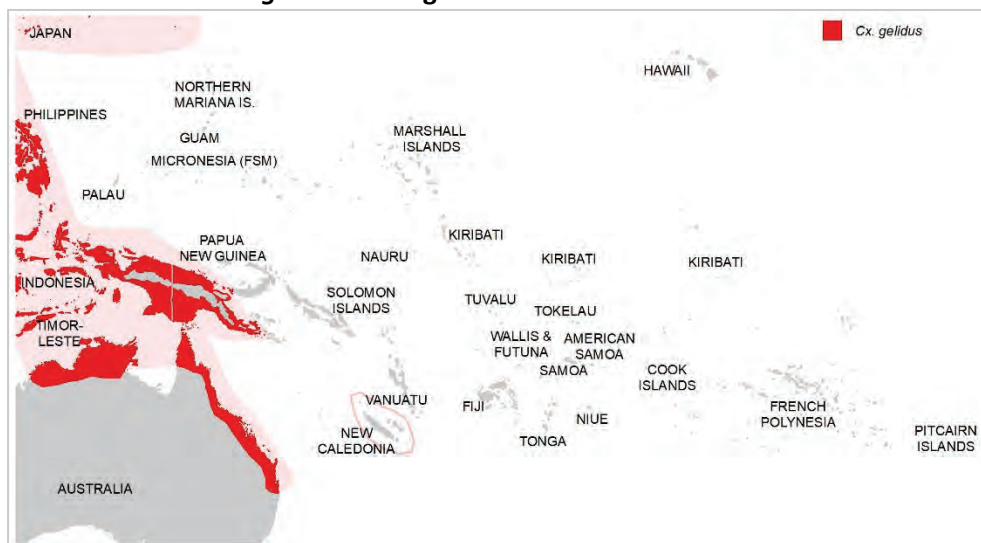


Photo source:
Stephen Doggett

Bionomics of *Culex gelidus*

Subgenus	<i>Culex</i>
Distribution	Widespread throughout southeast Asia and into New Guinea. Has been introduced to Australia. ^{24,27,31,81,206,225} Was recently detected in New Caledonia.
Aquatic habitats	Found in a variety of temporary and semi-permanent ground water habitats, usually with a high organic content, ranging from small ponds through to large wetlands and wastewater treatment ponds. Larvae have been found in natural and artificial containers. ³¹
Feeding times	The majority of feeding occurs at dusk and dawn (crepuscular) and continues throughout the night. ²⁴
Blood meal hosts	Prefers to feed on large domestic animals such as cows, horses and pigs. In the absence of domesticated livestock, they readily feed on humans. Other documented hosts include birds, chickens goats and deer. ²⁴ Is often a significant nuisance pest.
Feeding location	Either indoors or outdoors.
Resting habits	Outside of houses, likely in vegetation or animal sheds.
Flight range	Unknown.
Medical importance	Numerous arboviruses including Japanese encephalitis (secondary vector) ^{237,238} and Ross River virus (secondary vector). ²³⁹ One laboratory study suggested the potential of being a chikungunya vector, but the methodology bypassed any gut barriers to infection by using intrathoracic inoculation. ²⁴⁰

Culex quinquefasciatus Say 1823

Distribution of *Culex quinquefasciatus* throughout the Pacific Island countries

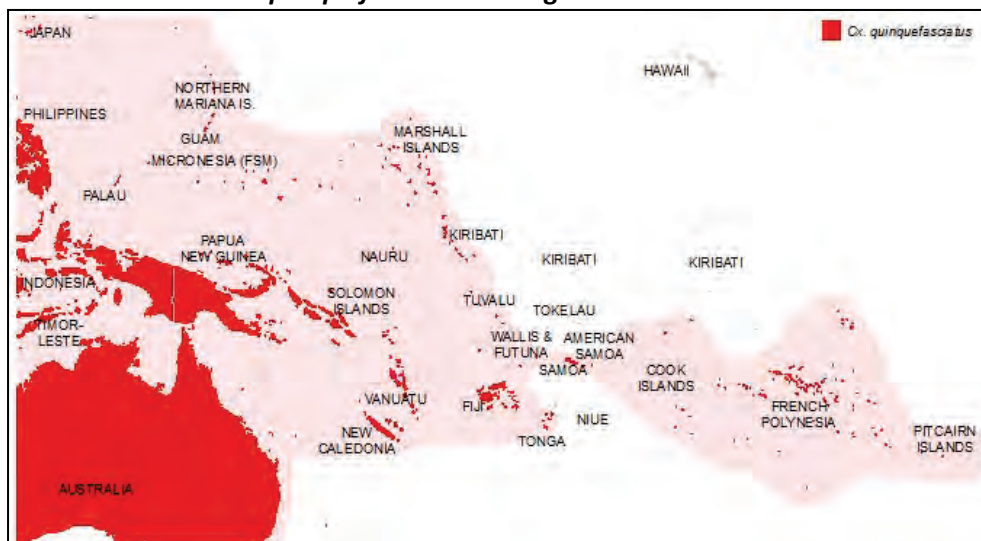


Photo source:
Stephen Doggett

Bionomics of *Culex quinquefasciatus*

Subgenus	<i>Culex</i>
Distribution	Widely distributed across globe and in the Pacific region ^{31,92,99,101,138,225} . Found more commonly in urban areas.
Aquatic habitats	Commonly found in organically rich water, including sewers, ground pools, ditches, poorly constructed septic tanks or cesspools. Will also use artificial containers ^{24,31} .
Feeding times	Feeds mostly at night, with a peak at midnight ⁹² .
Blood meal hosts	Predominantly anthropophilic, but recorded feeding on other warm-blooded vertebrates, including birds, domestic animals, and small mammals ^{24,34} . Is often a significant nuisance pest.
Feeding location	Readily feeds indoors or outdoors.
Resting habits	Indoors and outdoors.
Flight range	The flight range thought to be less than 2 km ¹¹² .
Medical importance	Numerous arboviruses, including Ross River virus (inefficient vector) ^{241,242} , Japanese encephalitis (competent vector in Asia, colonies of Australian populations indicate competence) ^{22,243} , <i>Wuchereria bancrofti</i> (secondary vector in Micronesia, inefficient vector in Melanesia and Polynesia) ^{64,80,202,225,244-247} , as well as <i>Dirofilaria immitis</i> (dog heart worm) ¹³⁵ . Has been observed to ingest chikungunya, but does not become infectious ^{248,249} .

Culex sitiens Wiedemann 1828

Distribution of *Culex sitiens* throughout the Pacific Islands

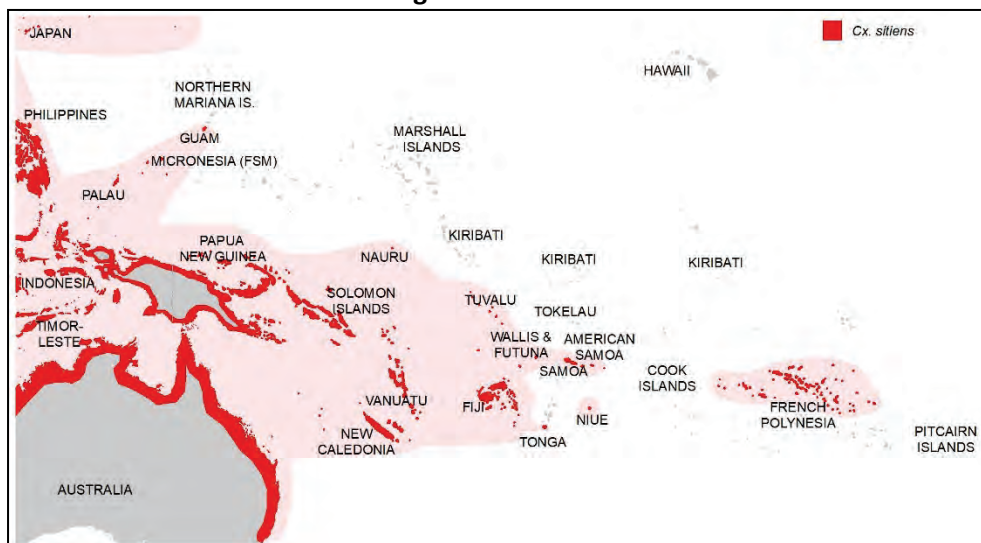


Photo source:
Stephen Doggett

Bionomics of *Culex sitiens*

Subgenus	<i>Culex</i>
Distribution	Africa, Asia, Australia and the Pacific. At the eastern limit of its range, <i>Culex sitiens</i> is found in Tonga (Tongatapu), the Samoan Islands, Niue and French Polynesia (Society, Tuamotu). ^{31,92,99,101,138,140,149,225}
Aquatic habitats	Associated with coastal habitats including inundated estuarine wetlands and saltmarsh pools as well as puddles, ditches, rock pools, ponds, crab holes. Water may be fresh or brackish. Noted in artificial containers. ^{24,31}
Feeding times	Bites throughout the night, with peak activity from 1900 to midnight. ^{24,92}
Blood meal hosts	Preferred host is birds, although it will bite humans. ³⁴ Although it is not usually a significant nuisance pest, high biting rates are observed. ²⁴
Feeding location	Outdoors.
Resting habits	Outdoors in vegetation.
Flight range	Up to 1 km. ^{31,250}
Medical importance	Some arboviruses including Japanese encephalitis virus (secondary vector). ^{22,251} For Ross River virus, has relatively low vector competence in the laboratory, and is considered to be an inefficient vector in the field. ^{229,230}

Culex tritaeniorhynchus Giles 1901

Distribution of *Culex tritaeniorhynchus* throughout the Pacific Islands

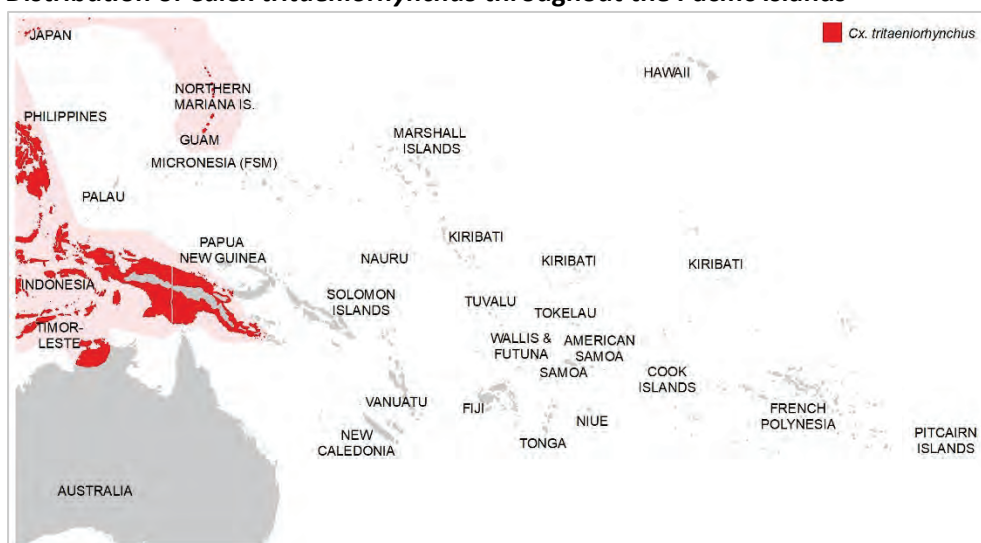


Photo source:
Walter Reed
Biosystematics Unit

Bionomics of *Culex tritaeniorhynchus*

Subgenus	<i>Culex</i>
Distribution	Eastern Africa through Asia ²²⁵ and in the Pacific in Papua New Guinea, ²⁴ Guam and the Northern Mariana Islands. ¹⁵⁵ Recently detected in northern Australia. ²⁵²
Aquatic habitats	Utilises a range of clean to polluted freshwater habitats including rice paddies, ground pools, wells, rock pools and stream margins. Also artificial containers. ^{24,145}
Feeding times	Feeds throughout the night from dusk to dawn.
Blood meal hosts	Almost exclusively on cows and water buffalo, feeds on pigs and humans when cows are not available. ^{24,253}
Feeding location	Strongly exophilic. ²⁴
Resting habits	Outdoors in vegetation or animal sheds. ¹⁴⁵
Flight range	The average flight range is 2 km. ¹¹²
Medical importance	Numerous arboviruses including Japanese encephalitis virus (primary vector in Asia). ²⁵⁴ Inhospitable to <i>Wuchereria bancrofti</i> . ^{225,255}

Mansonia uniformis (Theobald) 1901

Distribution of *Mansonia uniformis* throughout the Pacific Islands

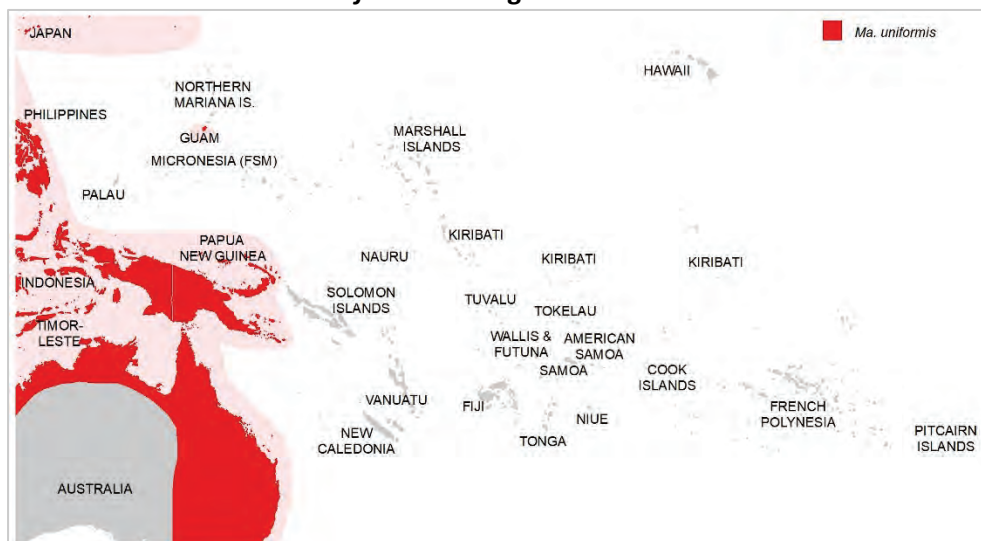


Photo source:
Stephen Doggett

Bionomics of *Mansonia uniformis*

Subgenus	<i>Mansonioides</i>
Distribution	Africa, Asia, Australia and in the Pacific in Papua New Guinea and Guam. ^{18,31,81,256}
Aquatic habitats	Permanent freshwater wetlands with abundant aquatic vegetation, particularly floating vegetation. The larvae have modified siphons that allow them to breathe by attaching to the tissues of aquatic plants. ³¹
Feeding times	Feeds throughout the night with peak biting between 1900 and 2000, just following sunset. ¹⁷⁰
Blood meal hosts	Primarily on cattle, but also frequently bites people and a range of other mammals. ²⁵⁷⁻²⁵⁹
Feeding location	Strongly exophagic, but will enter houses to feed. ²⁴
Resting habits	Rests outdoors in vegetation.
Flight range	The species is a strong flier, with flights detected of up to 4 km to larval habitats (average 1.5 km). ¹¹²
Medical importance	Numerous arboviruses including Japanese encephalitis virus (secondary vector) ²⁶⁰ and Ross River virus (primary vector). ²⁶¹ <i>Wuchereria bancrofti</i> (primary vector in Southeast Asia, secondary vector in New Guinea). ^{256,262,263}

Lists of mosquito species by country

American Samoa (12 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j
			<i>Finlaya</i>	-	-	<i>oceanicus</i>
				-	-	<i>tutuila</i>
			<i>Stegomyia</i>	<i>kochi</i>	-	<i>samoanus</i> ^F
				<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
	<i>scutellaris</i>	-		<i>polynesiensis</i> ^{d,c,z,F,r}		
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^j
				<i>sitiens</i>	-	<i>annulirostris</i> ^{R,J}
				-	-	<i>sitiens</i> ^j
	Mansoniini	<i>Coquillettidia</i>	<i>Coquillettidia</i>	-	-	<i>samoensis</i>
Toxorhynchitini	<i>Toxorhynchites</i>	<i>Toxorhynchites</i>	<i>splendens</i>	-	<i>amboinensis</i>	

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

Note: A suspected, but not confirmed, vector of lymphatic filariasis is *Aedes tutuila*.

References: 65,100,138,175,208,264,265

Cook Islands (5 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j
			<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>scutellaris</i>	-	<i>polynesiensis</i> ^{d,c,z,F,r}
				<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^j
				<i>sitiens</i>	-	<i>annulirostris</i> ^{R,J}

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

Note: *Aedes vexans* was initially detected in 2022 during routine surveillance.

References: 106,208,266

Federated States of Micronesia (25 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species					
Culicinae	Aedeomyiini	<i>Aedeomyia</i>	<i>Aedeomyia</i>	-	-	<i>catacticta</i>					
				<i>Aedimorphus</i>	-	-	<i>oakleyi</i>				
					-	-	<i>senyavinensis</i>				
					-	-	<i>trukensis</i>				
					<i>vexans</i>	-	<i>vexans</i> ^j				
			<i>Lorrainea</i>	-	-	<i>lamelliferus</i>					
				<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}					
				<i>maehleri</i>	-	<i>maehleri</i>					
				<i>Stegomyia</i>	-	-	<i>albopictus</i> ^{d,c,z,r,j}				
					<i>scutellaris</i>	-	<i>hakanssoni</i>				
	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>scutellaris</i>	-	-	<i>hensilli</i> ^{d,c,z}				
					-	-	<i>marshallensis</i> ^{d,f}				
					-	-	<i>palauensis</i>				
					-	-	<i>scutoscriptus</i>				
					-	-	<i>pipkini</i>				
					<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^{F,j}				
					<i>Culex</i>	<i>Culex</i>	<i>sitiens</i>	-	-	<i>annulirostris</i> ^{R,j}	
								-	-	<i>sitiens</i> ^j	
					Culicini	<i>Culex</i>	<i>Culiciomyia</i>	<i>fragilis</i>	-	-	<i>maplei</i>
									-	-	<i>nigropunctatus</i>
	-	-	<i>gossi</i>								
	<i>Lophoceraomyia</i>	<i>fraudatrix</i>	-	-			<i>kusaiensis</i>				
			-	-			<i>hilli</i>				
			-	-			<i>carolinensis</i>				
	<i>Lutzia</i>	<i>Metalutzia</i>			-	-	<i>fuscana</i>				
-					-	<i>vorax</i>					

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{102,155,157,165,166,267-270}

Fiji (27 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species				
Culicinae	Aedeomyiini	<i>Aedeomyia</i>	<i>Aedeomyia</i>	-	-	<i>catasticta</i>				
				<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j			
				<i>Finlaya</i>	-	-	<i>burnetti</i>			
					<i>kochi</i>	-	<i>fijiensis</i> ^f			
				<i>Levua</i>	-	-	<i>geoskusea</i>			
				Aedini	<i>Aedes</i>	<i>Ochlerotatus</i>	<i>emiphals</i>	-	<i>vigilax</i> ^{R,j}	
							<i>gilesia</i>	-	<i>mcdonaldi</i>	
							<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}	
							<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>albopictus</i> ^{d,C,z,r,j}
									-	<i>horrescens</i>
	-	<i>polynesiensis</i> ^{d,C,z,F,r}								
	-	<i>pseudoscutellaris</i> ^{d,F}								
	-	<i>rotumae</i> ^{d,f}								
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^{f,j}				
				<i>sitiens</i>	-	<i>annulirostris</i> ^{R,J}				
				<i>Oculeomyia</i>	-	<i>sitiens</i> ⁱ				
				-	-	<i>albinervis</i>				
				-	-	<i>fijiensis</i>				
				Mansoniini	<i>Coquillettidia</i>	<i>Coquillettidia</i>	<i>crassipes</i>	-	<i>crassipes</i>	
							<i>Polylepidomyia</i>	-	<i>rotumanus</i>	
				Sabethini	<i>Tripteroidea</i>	<i>Tripteroidea</i>	<i>nitidoventer</i>	-	<i>purpuratus</i>	
							-	-	<i>amboinensis</i>	
				Toxorhynchitini	<i>Toxorhynchites</i>	<i>Toxorhynchites</i>	<i>splendens</i>	-	<i>inornatus</i>	
	-	-	<i>splendens</i>							
	Uranotaeniini	<i>Uranotaenia</i>	<i>Pseudoficalbia</i>	-	-	<i>colocasiae</i>				
				-	-	<i>painei</i>				

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: 39,92,94,104,121,123,124,139,140,148,188,189,216,226,271-275

French Polynesia (14 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species		
Culicinae	Aedini	<i>Aedes</i>	<i>Ochlerotatus</i>	<i>buvirilia</i>	-	<i>edgari</i>		
			<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}		
				<i>scutellaris</i>	-	<i>polynesiensis</i> ^{d,c,z,F,r}		
			Culicini	<i>Culex</i>	<i>Culex</i>	<i>atriceps</i>	-	<i>atriceps</i>
						<i>atriceps</i>	-	<i>kesseli</i>
						<i>atriceps</i>	-	<i>marquesensis</i>
	<i>pipiens</i>	<i>pipiens</i>				<i>quinquefasciatus</i> ^j		
	<i>pipiens</i>	<i>pipiens</i>				<i>annulirostris</i> ^{R,j}		
	<i>pipiens</i>	<i>pipiens</i>				<i>roseni</i>		
	<i>sitiens</i>	-				<i>sechani</i>		
	<i>sitiens</i>	-				<i>sitiens</i> ^j		
	Sabethini	<i>Wyeomyia</i>	<i>Wyeomyia</i>	-	-	<i>mitchellii</i>		
	Toxorhynchitini	<i>Toxorhynchites</i>	<i>Toxorhynchites</i>	<i>splendens</i>	-	<i>amboinensis</i>		

Note: *Aedes vexans* is most likely not found in French Polynesia. Whilst it has been reported once in 2004²⁰⁹, the observation seems anecdotal, and this species has not been detected again despite extensive surveys.

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: 98,99,180,182,209,226,276-278

Guam (29 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species	
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Anopheles</i>	<i>barbirostris</i>	<i>barbirostris</i>	<i>barbirostris</i> ^{m,f}	
						<i>campestris</i> ^{m,f}	
				<i>hyrcanus</i>	-	<i>lesteri</i> ^m	
				<i>umbrosus</i>	-	<i>baezai</i>	
						<i>indefinitus</i> ^M	
					<i>litoralis</i> ^m		
					<i>vagus</i> ^m		
					<i>subpictus</i>	<i>subpictus</i> ^{M,f}	
					<i>tessellatus</i>	-	<i>tessellatus</i>
							<i>catasticta</i>
Culicinae	Aedomyiini	<i>Aedomyia</i>	<i>Aedomyia</i>	-	-	<i>oakleyi</i>	
						<i>trimaculatus</i>	
	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	-	-	<i>vexans</i> ^j	
						<i>pandani</i>	
						<i>rotanus</i>	
						<i>saipanensis</i>	
			<i>Stegomyia</i>			<i>albopictus</i> ^{D,C,Z,r,j}	
						<i>guamensis</i>	
						<i>subalbatus</i> ^j	
						<i>fuscocephala</i> ^j	
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	-	-	<i>vagens</i>
							<i>quinquefasciatus</i> ^{F,j}
							<i>annulirostris</i> ^{R,j}
							<i>litoralis</i>
<i>sitiens</i>						<i>sitiens</i> ^j	
						<i>tritaeniorhynchus</i> ^j	
						<i>fuscana</i>	
						<i>uniformis</i> ^{F,R,j}	
Mansoniini	<i>Mansonia</i>	<i>Mansonioides</i>	-	-	<i>uniformis</i> ^{F,R,j}		
Sabethini	<i>Wyeomyia</i>	<i>Wyeomyia</i>	-	-	<i>mitchellii</i>		

Legend for vector status: M or m: malaria, D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

For *Culex annulirostris*, the subspecies found in Guam is *Culex annulirostris marianae*.

References: ^{18,146,279-283}

Guam – species not detected since 2010 and likely eliminated

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i>

References: ¹⁴⁶

Kiribati (6 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species	
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^J	
			<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}	
				<i>scutellaris</i>	-	<i>albopictus</i> ^{d,c,z,r,j}	
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	<i>pipiens</i>	-	<i>quinquefasciatus</i> ^{F,j}
				<i>sitiens</i>	-	<i>annulirostris</i> ^{R,j}	

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{39,96,121,208}

Marshall Islands (5 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species	
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^J	
			<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}	
				<i>scutellaris</i>	-	<i>albopictus</i> ^{d,c,z,r,j}	
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	<i>pipiens</i>	-	<i>quinquefasciatus</i> ^{F,j}

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{17,39,97,208}

Nauru (5 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
				<i>scutellaris</i>	-	<i>albopictus</i> ^{d,c,z,r,j}
				<i>pipiens</i>	<i>pipiens</i>	-
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>sitiens</i>	-	<i>annulirostris</i> ^{R,j}

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{39,121}

New Caledonia (21 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species	
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Anopheles</i>	<i>bancroftii</i>	<i>bancroftii</i>	<i>bancroftii</i> A ^{m,f,r}	
				<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j
	Aedini	<i>Aedes</i>		<i>Mucidus</i>	<i>mucidus</i>	-	<i>alternans</i>
				<i>Ochlerotatus</i>	<i>emiphals</i>	-	<i>vigilax</i> ^{F,R,j}
				<i>Rampamyia</i>	<i>notoscriptus</i>	-	<i>notoscriptus</i> ^r
				<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
						-	<i>iyengari</i>
					<i>pipiens</i>	<i>pipiens</i>	<i>australicus</i>
	Culicinae	Culicini	<i>Culex</i>		<i>Culex</i>		<i>quinquefasciatus</i> ^{f,j}
							<i>annulirostris</i> ^{R,j}
					<i>sitiens</i>	-	<i>gelidus</i> ^{r,j}
						-	<i>sitiens</i> ^j
						-	<i>cheesmanae</i>
		<i>Neoculex</i>	<i>Pseudomelanoconia</i>	-	<i>dumbletoni</i>		
				-	<i>gaufini</i>		
				-	<i>millironi</i>		
			<i>Oculeomyia</i>	-	-	<i>starckeae</i>	
					<i>bitaeniorhynchus</i>	<i>bitaeniorhynchus</i> ^j	
	Mansoniini	<i>Coquillettida</i>	<i>Coquillettida</i>	-	-	<i>xanthogaster</i> ^r	
	Sabethini	<i>Tripteroides</i>	<i>Polylepidomyia</i>	-	-	<i>caledonicus</i>	
						<i>melanesiensis</i>	

Legend for vector status: M or m: malaria, D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{17,39,121,208,222,284}

New Caledonia – species likely eliminated

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>scutellaris</i>

Aedes scutellaris was detected in New Caledonia for the first time in 2016. An eradication plan was put in place, and it has not been detected since December 2017, and is therefore considered eliminated.

Niue (4 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
				<i>scutellaris</i>	-	<i>cooki</i> ^{d,f}
	Culicini	<i>Culex</i>	<i>Culex</i>			<i>polynesiensis</i> ^{d,c,z,F,r}
				<i>sitiens</i>	-	<i>sitiens</i> ^j

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{39,121}

Northern Mariana Islands (16 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species			
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Cellia</i>	-	-	<i>indefinitus</i> ^M			
			<i>Aedimorphus</i>	-	-	<i>oakleyi</i>			
	<i>vexans</i>	-		-	<i>vexans</i> ^J				
	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>pandani</i>	-	-	<i>agrihanensis</i>		
					-	-	<i>neopandani</i>		
					-	-	<i>pandani</i>		
					-	-	<i>rotanus</i>		
			Culicinae	Culicini	<i>Culex</i>	<i>Culex</i>	<i>sitiens</i>	-	<i>saipanensis</i>
								-	<i>albopictus</i> ^{D,C,Z,r,j}
								-	<i>guamensis</i>
-								<i>quinquefasciatus</i> ^{F,j}	
Culicinae	Culicini	<i>Lutzia</i>	<i>Metalutzia</i>	-	-	<i>annulirostris</i> ^{R,j}			
						<i>litoralis</i>			
						<i>tritaeniorhynchus</i> ^J			
Culicinae	Culicini	<i>Lutzia</i>	<i>Metalutzia</i>	-	-	<i>fuscana</i>			
						<i>vorax</i>			

Legend for vector status: M or m: malaria, D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

For *Culex annulirostris*, the subspecies found in the Northern Mariana Islands is *Culex annulirostris marianae*.

References: 18,39,155,283,285,286

Northern Mariana Islands – species not detected since 2010 and likely eliminated

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i>

Palau (21 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species					
Culicinae	Aedeomyiini	<i>Aedeomyia</i>	<i>Aedeomyia</i>	-	-	<i>catasticta</i>					
			<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j					
			<i>Downsiomyia</i>	<i>niveus</i>	-	<i>dorseyi</i>					
			<i>Finlaya</i>	-	-	<i>gressitti</i>					
						<i>hui</i>					
	Aedini	<i>Aedes</i>		<i>Lorrainea</i>	-	-	<i>lamelliferus</i>				
					<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}				
							<i>albopictus</i> ^{d,c,z,r,j}				
							<i>dybasi</i>				
							<i>hensilli</i> ^{d,c,z}				
	Culicini		<i>Culex</i>	<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>palauensis</i>				
							<i>scutellaris</i> ^d				
							<i>Verrallina</i>	<i>Verrallina</i>	-	-	<i>pipkini</i>
									<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^{F,j}
									<i>sitiens</i>	-	<i>annulirostris</i> ^{R,j}
											<i>sitiens</i> ^j
									<i>Culiciomyia</i>	<i>fragilis</i>	-
								<i>Oculeomyia</i>	-	<i>bitaeniorhynchus</i>	<i>bitaeniorhynchus</i> ^j
								<i>Lutzia</i>	<i>Metalutzia</i>	-	<i>fusca</i>
											<i>crassipes</i>
Mansoniini	<i>Coquillettidia</i>	<i>Coquillettidia</i>	<i>crassipes</i>	-	<i>crassipes</i>						

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{39,155,163}

Papua New Guinea (284 species)

Sub-family	Tribe	Genus	Subgenus	Group	Complex	Species
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Anopheles</i>	<i>bancroftii</i>	-	<i>pseudobarbirostris</i>
					<i>bancroftii</i>	<i>bancroftii</i> A ^{m,f,r}
						<i>bancroftii</i> B ^{m,f}
						<i>bancroftii</i> C ^{m,f}
						<i>bancroftii</i> D ^{m,f}
					-	<i>papuensis</i>
			<i>Cellia</i>	-	<i>hilli</i>	
				-	<i>incognitus</i>	
				-	<i>karwari</i> ^m	
				-	<i>meraukensis</i>	
				-	<i>novaguinensis</i>	
				<i>longirostris</i>	<i>longirostris</i> A ^m	
					<i>longirostris</i> B ^m	
					-	<i>longirostris</i> C1 ^m
					-	<i>longirostris</i> C2 ^m
					<i>longirostris</i> D ^m	
					<i>longirostris</i> E ^m	
					<i>longirostris</i> F ^m	
			<i>longirostris</i> G ^m			
			<i>annulipes</i>	<i>annulipes</i>	<i>annulipes</i> ^m	
				<i>annulipes</i>	<i>annulipes</i> ^f	
				-	<i>clowi</i>	
				-	<i>punctulatus</i> ^{M,F}	
				<i>punctulatus</i>	<i>punctulatus</i>	<i>sp. nr. punctulatus</i>
<i>punctulatus</i>	<i>farauti</i> 4 ^M					
<i>punctulatus</i>	<i>farauti</i> 5					
<i>punctulatus</i>	<i>farauti</i> 8 ^m					
<i>farauti</i>	<i>farauti</i>	<i>farauti</i> s.s. ^{M,F}				
	<i>farauti</i>	<i>hinesorum</i> ^{M,F}				
	<i>farauti</i>	<i>oreios</i> ^m				
	<i>farauti</i>	<i>torresiensis</i>				
<i>koliensis</i>	<i>koliensis</i>	<i>koliensis</i> 1 ^{M,F}				
	<i>koliensis</i>	<i>koliensis</i> 3 ^{M,F}				
<i>Bironella</i>	<i>Bironella</i>	-	<i>gracilis</i>			
		-	<i>simmondsi</i>			
		-	<i>hollandi</i>			
		-	<i>obscura</i>			
		-	<i>travestita</i>			
		-	<i>papuae</i>			
<i>Neobironella</i>	<i>Neobironella</i>	-	-			
		-	-			
Culicinae	Aedeomyiini	<i>Aedeomyia</i>	<i>Aedeomyia</i>	-	-	<i>catasticta</i>

Sub-family	Tribe	Genus	Subgenus	Group	Complex	Species	
Culicinae	Aedini	<i>Aedes</i>		-	-	<i>lowisii</i>	
			<i>Aedimorphus</i>		<i>alboscutecllatus</i>	-	<i>alboscutecllatus</i>
					<i>vexans</i>	-	<i>vexans</i> ^j
			<i>Downsiomyia</i>		<i>niveus</i>	-	<i>niveus</i> <i>shehzadae</i>
			<i>Edwardsaedes</i>		-	-	<i>imprimens</i> ^r
					-	-	<i>gani</i>
					-	-	<i>kochi</i> ^{f,r}
			<i>Finlaya</i>		<i>kochi</i>	-	<i>wallacei</i>
						<i>bougainvillensis</i>	<i>bougainvillensis</i> <i>hollingsheadi</i> <i>neogeorgianus</i>
			<i>Geoskusea</i>		-	-	<i>fimbripes</i>
			<i>Huaedes</i>		-	-	<i>wauensis</i>
			<i>Leptosomatomyia</i>		-	-	<i>aurimargo</i>
			<i>Lorrainea</i>		-	-	<i>dasyorrhus</i>
			<i>Macleaya</i>		<i>macleaya</i>	-	<i>littlechildi</i> <i>tremulus</i>
						<i>mucidus</i>	<i>alternans</i>
			<i>Mucidus</i>		<i>pardomyia</i>	-	<i>aurantius</i> <i>nigrescens</i>
						-	<i>inexpectatus</i> <i>normanensis</i> ^r <i>vittiger</i>
			<i>Ochlerotatus</i>		-	-	<i>vigilax</i> ^{R,j}
						<i>emiphals</i>	<i>mcdonaldi</i> <i>purpuraceus</i>
						<i>gilesia</i>	<i>purpuraceus</i> <i>anggiensis</i> <i>argenteitarsis</i> <i>clintoni</i>
			<i>Patmarksia</i>		-	-	<i>dobodurus</i> <i>hollandius</i> <i>novalbitarsis</i> <i>papuensis</i>
			<i>Phagomyia</i>		-	-	<i>plumiferus</i>
			<i>Pseudoskusea</i>		-	-	<i>culiciformis</i>
			<i>Rampamyia</i>		-	-	<i>albilabris</i>
						<i>notoscriptus</i>	<i>notoscriptus</i> ^r
			<i>Rhinoskusea</i>		-	-	<i>longirostris</i>
			<i>Scutomomyia</i>		-	-	<i>albolineatus</i>
						<i>aegypti</i>	<i>aegypti</i> ^{D,C,Z,r}
						<i>annandalei</i>	<i>annandalei</i> <i>albopictus</i> ^{d,c,z,r,j} <i>gurneyi</i>
			<i>Stegomyia</i>			<i>scutellaris</i>	<i>hebrideus</i> ^d <i>quasiscutellaris</i> <i>scutellaris</i> ^d <i>varuae</i>
							<i>candidoscutellum</i> <i>gracilelineatus</i> <i>keefeii</i>
			<i>Uncertain placement</i>		-	-	<i>quasirubithorax</i> <i>rubiginosus</i> <i>stanleyi</i> <i>tsiliensis</i>

Sub-family	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	Armigeres	Armigeres	-	-	<i>breinli</i>
						<i>denbesteni</i>
						<i>lacuum</i>
						<i>malayi</i>
						<i>milnensis</i>
						<i>obturbans</i>
		Verrallina	Verrallina	-	-	<i>neomacrodixoa</i>
						<i>panayensis</i>
						<i>carmenti</i> ^r
						<i>cuccioi</i>
						<i>embiensis</i>
						<i>foliformis</i>
						<i>funerea</i> ^R
						<i>killertonis</i>
						<i>leilae</i>
	Verrallina	Verrallina	-	-	<i>lineata</i> ^r	
					<i>mccormicki</i>	
					<i>milnensis</i>	
					<i>multifolium</i>	
					<i>obsoleta</i>	
					<i>parasimilis</i>	
	Culicini	Culex	Culex	-	-	<i>quadrifolium</i>
						<i>quadrispinata</i>
						<i>reesi</i>
						<i>sentania</i>
						<i>similis</i>
						<i>trispinata</i>
<i>vanapa</i>						
<i>variabilis</i>						
<i>bicki</i>						
<i>binigrolineatus</i>						
<i>axillicolus</i>						
<i>belkini</i>						
<i>bougainvillensis</i>						
<i>pallidiceps</i>						
<i>perkinsi</i>						
<i>miraculosus</i>						
<i>pervigilans</i>						
<i>quinqüefasciatus</i> ^{f,j}						
<i>alis</i>						
<i>annulirostris</i> ^{f,R,j}						
<i>edwardsi</i>						
<i>gelidus</i> ^{r,j}						
<i>mimulus</i>						
<i>palpalis</i> ^{r,j}						
<i>perplexus</i>						
<i>sitiens</i> ^l						
<i>solitarius</i>						
<i>tritaeniorhynchus</i> ^l						
<i>vicinus</i>						
<i>whitmorei</i>						

Sub-family	Tribe	Genus	Subgenus	Group	Complex	Species	
Culicinae	Culicini	<i>Culex</i>	<i>Culiciomyia</i>	<i>fragilis</i>	-	<i>fragilis</i>	
					-	<i>nailoni</i>	
					-	<i>papuensis</i>	
				-	-	<i>pullus</i>	
				-	<i>nebulosus</i>	<i>ruthae</i>	
				-	<i>shebbearei</i>	<i>bailyi</i>	
				<i>Eumelanomyia</i>	<i>mochthogenes</i>	-	<i>cataractarum</i>
						-	<i>manusensis</i>
					<i>protomelanoconion</i>	-	<i>brevipalpis</i>
						-	<i>bergi</i>
					-	<i>buxtoni</i>	<i>lairdi</i>
					-	<i>christiani</i>	<i>christiani</i>
			<i>Lophoceraomyia</i>	<i>fraudatrix</i>	-	<i>gressitti</i>	
					-	<i>minjensis</i>	
					-	<i>pseudornatus</i>	
					-	<i>cottlei</i>	
					-	<i>cottlei</i>	
					-	<i>atracus</i>	
				<i>fraudatrix</i>	-	<i>collessi</i>	
					-	<i>fraudatrix</i>	
					-	<i>insularis</i>	
					-	<i>kaviengensis</i>	
					-	<i>rajaneae</i>	
					-	<i>schilfgaardei</i>	
			<i>Lophoceraomyia</i>	<i>fraudatrix</i>	-	<i>submarginalis</i>	
					-	<i>hilli</i>	
					-	<i>lakei</i>	
					-	<i>hurlbuti</i>	
					-	<i>hurlbuti</i>	
					-	<i>kowiroensis</i>	
				<i>fraudatrix</i>	-	<i>muruae</i>	
					-	<i>versabilis</i>	
					-	<i>wamanguae</i>	
-	<i>minutissimus</i>						
-	<i>cylindricus</i>						
-	<i>ornatus</i>						
<i>Lophoceraomyia</i>	<i>fraudatrix</i>	-	<i>gagnei</i>				
		-	<i>crowei</i>				
		-	<i>petersi</i>				
		-	<i>shanahani</i>				
		-	<i>steffani</i>				
		-	<i>castaneus</i>				
	<i>fraudatrix</i>	-	<i>pseudorubithoracis</i>				
		-	<i>pseudorubithoracis</i>				
		-	<i>sedlacekae</i>				
		-	<i>durhami</i>				
		-	<i>walukasi</i>				
		-	<i>solomonis</i>				
<i>Lophoceraomyia</i>	<i>fraudatrix</i>	-	<i>cubitatus</i>				
		-	<i>variatus</i>				
		<i>mammilifer</i>	-	<i>bolii</i>			
			-	<i>digoelensis</i>			
			-	<i>kuhnsi</i>			
			-	<i>singuawaensis</i>			
	-		<i>squamosus</i>				
	-		<i>bitaeniorhynchus</i>				
	<i>Oculeomyia</i>	-	<i>bitaeniorhynchus</i> ^{f,j}				
		-	<i>sinensis</i>				
		-	<i>halifaxii</i>				
		-	<i>minima</i>				
-		<i>elegans</i>					
-		<i>gurneyi</i>					
Filcalbiini	<i>Lutzia</i>	-	<i>chamberlaini</i>				
		-	<i>cairnsensis</i>				
	<i>Ficalbia</i>	-	<i>quasisanguinae</i>				
		-	<i>solomonis</i>				
	<i>Mimomyia</i>	-	<i>spoliata</i>				
		-					
Hodgesiini	<i>Hodgesia</i>	-					
		-					

Sub-family	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Mansoniini	<i>Coquillettidia</i>	<i>Coquillettidia</i>	-	-	<i>fuscopteron</i>
						<i>giblini</i>
						<i>linealis</i> ^R
						<i>memorans</i>
						<i>ochracea</i>
		<i>Mansonia</i>	<i>Mansonioides</i>	-	-	<i>xanthogaster</i> ^r
						<i>crassipes</i>
						<i>annulifera</i>
						<i>indiana</i>
						<i>melanesiensis</i>
	Orthopodomyiini	<i>Orthopodomyia</i>	-	-	-	<i>papuensis</i>
						<i>genurostris</i>
						<i>leei</i>
						<i>solomonis</i>
						<i>papuensis</i>
						<i>altivallis</i>
						<i>argenteiventris</i>
						<i>microlepis</i>
						<i>perplexus</i>
						<i>solomonis</i>
						<i>standfasti</i>
						<i>bisquamatus</i>
						<i>brevirhynchus</i>
						<i>concinnus</i>
						<i>confusus</i>
						<i>filipes</i>
						Sabethini
<i>leei</i>						
<i>longipalpatus</i>						
<i>mathesoni</i>						
<i>stonei</i>						
<i>torokinae</i>						
<i>vanleeuweni</i>						
<i>alboscuteallatus</i>						
<i>bimaculipes</i>						
<i>binotatus</i>						
<i>brevipalpis</i>						
Toxorhynchitini	<i>Toxorhynchites</i>	<i>Tripteroides</i>	<i>nitidoventer</i>	-	<i>elegans</i>	
					<i>littlechildi</i>	
					<i>lorengai</i>	
					<i>magnesianus</i>	
					<i>nissanensis</i>	
					<i>novohanoverae</i>	
					<i>quasiornatus</i>	
					<i>splendens</i>	
					<i>amboinensis</i>	
					<i>inornatus</i>	
<i>nepenthicola</i>						
<i>speciosus</i>						
<i>splendens</i>						

Sub-family	Tribe	Genus	Subgenus	Group	Complex	Species		
Culicinae	Uranotaeniini	Uranotaenia	<i>Pseudoficalbia</i>	-	-	<i>atra</i>		
						<i>quadrimaculata</i>		
						<i>diagonalis</i>		
						<i>hirsutifemora</i>		
						<i>obscura</i>		
						<i>albescens</i>		
			<i>Uranotaenia</i>	-	-	<i>albosternopleura</i>		
						<i>amiensis</i>		
						<i>antennalis</i>		
						<i>argyrotarsis</i>		
						<i>barnesi</i>		
						<i>civinskii</i>		
						<i>lateralis</i>		
		<i>Uranotaenia</i>	-	-	<i>moresbyensis</i>			
					<i>neotibialis</i>			
					<i>nivipes</i>			
					<i>novaguinensis</i>			
					<i>paralateralis</i>			
					<i>paranovaguinensis</i>			
								<i>setosa</i>
								<i>sexaueri</i>
								<i>solomonis</i>
								<i>tibialis</i>
<i>tibioclada</i>								
<i>wysockii</i>								

Legend for vector status: M or m: malaria, D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: 29,38,39,81,121,123,264

Pitcairn Islands (3 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
				<i>scutellaris</i>	-	<i>polynesiensis</i> ^{d,c,z,F,r}
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^j

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{39,226}

Samoa (14 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species			
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j			
				<i>Finlaya</i>	-	-	<i>oceanicus</i> <i>tutuillae</i>		
			<i>Stegomyia</i>	<i>kochi</i>	-	<i>samoanus</i> ^F			
				<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}			
				<i>albopictus</i> ^{d,c,z,r,j}	-	<i>polynesiensis</i> ^{d,c,z,F,r}			
				<i>scutellaris</i>	-	<i>upolensis</i> ^f			
				<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^j			
				<i>sitiens</i>	-	<i>annulirostris</i> ^{R,J} <i>sitiens</i> ^j			
			Culicini	<i>Culex</i>	<i>Culex</i>	<i>Oculeomyia</i>	-	-	<i>samoensis</i>
			Mansoniini	<i>Coquillettidia</i>	<i>Coquillettidia</i>	-	-	<i>samoensis</i>	
	Toxorhynchitini	<i>Toxorhynchites</i>	<i>Toxorhynchites</i>	<i>splendens</i>	-	<i>amboinensis</i>			

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

Note: A suspected, but not confirmed, vector of lymphatic filariasis is *Aedes tutuillae*.

References: ^{39,121,138,201,208,264}

Solomon Islands (115 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species								
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Cellia</i>	<i>lungae</i>	-	<i>lungae</i> s.s.								
						<i>nataliae</i>								
						<i>solomonis</i>								
				<i>punctulatus</i>	<i>farauti</i>	<i>punctulatus</i> ^{m,f}								
						<i>rennellensis</i>								
						<i>farauti</i> s.s. ^{M,F}								
						<i>hinesorum</i> ^{m,f}								
						<i>irenicus</i>								
						<i>hollandi</i>								
						<i>catasticta</i>								
				<i>Aedeomyia</i>	<i>Aedeomyia</i>	<i>Aedimorphus</i>	<i>alboscutellarus</i>	-	<i>alboscutellarus</i>					
							<i>vexans</i>	-	<i>vexans</i> ^l					
						<i>Christophersiomyia</i>	-	-	<i>chionodes</i>					
						<i>Edwardsaedes</i>	-	-	<i>imprimens</i> ^r					
						<i>Finlaya</i>	<i>Finlaya</i>	<i>kochi</i>			<i>horotoi</i>			
											<i>bougainvillensis</i>			
											<i>hollingsheadi</i>			
											<i>maffii</i>			
											<i>neogeorgiansus</i>			
<i>schlosseri</i>														
<i>franclemonti</i>	<i>franclemonti</i>													
<i>knighti</i>	<i>knighti</i>													
<i>solomonis</i>	<i>fuscipalpis</i>													
	<i>fuscitarsis</i>													
<i>solomonis</i>	<i>solomonis</i>													
Culicinae	Aedini	<i>Aedes</i>				<i>becki</i>								
						<i>daggyi</i>								
						<i>Geoskusea</i>	<i>Geoskusea</i>	-	-	<i>fimbripes</i>				
										<i>longiforceps</i>				
										<i>perryi</i>				
						<i>Lorrainea</i>	-	-	<i>dasyorrhus</i>					
						<i>Mucidus</i>	<i>pardomyia</i>	-	<i>nigrescens</i>					
						<i>Ochlerotatus</i>	<i>emiphals</i>	-	<i>vigilax</i> ^{R,j}					
							<i>gilesia</i>	-	<i>mcdonaldi</i>					
						<i>Patmarksia</i>	-	-	<i>argyronotum</i>					
						<i>Rampamyia</i>	<i>Rampamyia</i>	-	-	<i>buxtoni</i>				
										<i>albilabris</i>				
						<i>Scutomya</i>	<i>Scutomya</i>	-	-	<i>notoscriptus</i>	<i>notoscriptus</i> ^r			
										<i>albolineatus</i>				
						<i>Stegomyia</i>	<i>Stegomyia</i>	<i>aegypti</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}			
											<i>robinsoni</i>			
											<i>edwardsi</i>	-	<i>tulagiensis</i>	
											<i>scutellaris</i>	<i>scutellaris</i>	-	<i>albopictus</i> ^{d,c,z,r,j}
														<i>gurneyi</i>
						<i>hebrideus</i> ^d								
<i>hoguei</i>														
<i>quasiscutellaris</i>														
<i>varuae</i>														
<i>Uncertain placement</i>	<i>Uncertain placement</i>	-	-	<i>roai</i>										
				<i>rubilabris</i>										

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species				
Culicinae	Aedini	<i>Armigeres</i>	<i>Armigeres</i>	-	-	<i>breinli</i>				
				<i>milnensis</i>						
		<i>Neomacleaya</i>	-	-	<i>neomacrodixoa</i>					
			<i>carmenti</i> ^r							
		<i>Verrallina</i>	<i>Verrallina</i>	-	-	<i>cuccioi</i>				
				<i>lineata</i> ^r						
		Culicini	<i>Culex</i>	<i>Acallyntrum</i>	<i>Acallyntrum</i>	<i>perkinsi</i>	-	<i>mccormicki</i>		
							<i>belkini</i>			
							<i>bougainvillensis</i>			
							<i>pallidiceps</i>			
	<i>perkinsi</i>									
	<i>quinquefasciatus</i> ^{f,j}									
	<i>Culex</i>			<i>Culex</i>	<i>sitiens</i>	-	<i>annulirostris</i> ^{R,j}			
						<i>omani</i>				
						<i>sitiens</i> ^j				
	<i>Culiciomyia</i>			<i>Culiciomyia</i>	<i>fragilis</i>	-	<i>whittingtoni</i>			
						<i>fragilis</i>				
						<i>papuensis</i>				
						<i>pullus</i>				
						<i>Eumelanomyia</i>	<i>Eumelanomyia</i>	<i>mochthogenes</i>	-	<i>fragilis</i>
									<i>femineus</i>	
									<i>bergi</i>	
									<i>lafooni</i>	
						<i>Lophoceraomyia</i>	<i>Lophoceraomyia</i>	<i>fraudatrix</i>	-	<i>oweni</i>
	<i>bergi</i>									
	<i>winkleri</i>									
	<i>buxtoni</i>									
	<i>lairdi</i>									
	<i>atracus</i>									
	<i>fraudatrix</i>									
	<i>Neoculex</i>	<i>Neoculex</i>	<i>crassistylus</i>	-	<i>hurlbuti</i>					
				<i>hurlbuti</i>						
<i>perryi</i>										
<i>becki</i>										
<i>solomonis</i>										
<i>Oculeomyia</i>	<i>Oculeomyia</i>		-	<i>solomonis</i>						
			<i>walukasi</i>							
<i>Lutzia</i>	<i>Lutzia</i>	<i>Metalutzia</i>	-	<i>leonardi</i>						
			<i>halifaxii</i>							
Filcalbiini	<i>Mimomyia</i>	<i>Etorleptiomyia</i>	-	<i>bougainvillensis</i>						
			<i>elegans</i>							
Hodgesiini	<i>Hodgesia</i>		-	<i>solomonis</i>						
			<i>solomonis</i>							
Mansoniini	<i>Coquillettidia</i>	<i>Coquillettidia</i>	<i>crassipes</i>	<i>lutea</i>						
			-	<i>melanesiensis</i>						
			-	<i>solomonis</i>						
Sabethini	<i>Tripteroides</i>	<i>Tripteroides</i>	-	<i>coheni</i>						
			<i>floridensis</i>							
			<i>melanesiensis</i>							
			<i>solomonis</i>							
		<i>Rachisoura</i>	<i>Rachisoura</i>	-	<i>mathesoni</i>					
				<i>stonei</i>						
				<i>torokinae</i>						
<i>Tripteroides</i>	<i>Tripteroides</i>	<i>nitidoventer</i>	-	<i>binotatus</i>						
			<i>bonneit</i>							
			<i>distigma</i>							
					<i>lipovskyi</i>					

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Uranotaeniini	<i>Uranotaenia</i>	<i>Pseudoficalbia</i>	-	-	<i>atra</i>
						<i>hirsutifemora</i>
						<i>painei</i>
						<i>quadrimaculata</i>
						<i>barnesi</i>
			<i>Uranotaenia</i>	-	-	<i>civinskii</i>
						<i>lateralis</i>
						<i>sexaueri</i>
						<i>solomonis</i>
						<i>wysockii</i>

Legend for vector status: M or m: malaria, D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: 33,40,46,65,71,90,123,149,150,168,287,288

Solomon Islands – species not detected since 2010 and likely eliminated

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Anophelinae	Anophelini	<i>Anopheles</i>	<i>Cellia</i>	<i>punctulatus</i>	<i>koliensis</i>	<i>koliensis</i>

References: 48,287,288

Tokelau (3 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j
			<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
				<i>scutellaris</i>	-	<i>polynesiensis</i> ^{d,c,z,F,r}

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: 96,107,121,208,289,290

Tonga (13 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j
			<i>Finlaya</i>	-	-	<i>oceanicus</i> ^f
				<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}
						<i>albopictus</i> ^{d,c,z,r,j}
						<i>cooki</i> ^{d,f}
			<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>horrescens</i>
						<i>kesseli</i> ^{d,f}
					<i>tongae</i>	<i>tabu</i> ^{d,F}
						<i>tongae s.s.</i> ^{d,F}
					<i>pipiens</i>	<i>pipiens</i>
Culicini	<i>Culex</i>	<i>Culex</i>		<i>sitiens</i>	-	<i>annulirostris</i> ^{R,j}
						<i>sitiens</i> ^j
			<i>Oculeomyia</i>	-	-	<i>albinervis</i>

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{101,105,123,136-138}

Tuvalu (6 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species	
Culicinae	Aedini	<i>Aedes</i>	<i>Aedimorphus</i>	<i>vexans</i>	-	<i>vexans</i> ^j	
			<i>Stegomyia</i>	<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}	
				<i>scutellaris</i>	-	<i>polynesiensis</i> ^{d,c,z,F,r}	
	Culicini	<i>Culex</i>	<i>Culex</i>		<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^j
					<i>sitiens</i>	-	<i>annulirostris</i> ^{R,j}
						<i>sitiens</i> ^j	

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: ^{39,121}

Vanuatu (22 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species			
Culicinae	Anophelini	<i>Anopheles</i>	<i>Cellia</i>	<i>punctulatus</i>	<i>farauti</i>	<i>farauti</i> s.s. ^{M,F}			
				<i>vexans</i>	-	<i>vexans</i> ^j			
				-	-	<i>daggi</i>			
				<i>emiphals</i>	-	<i>vigilax</i> ^{R,j}			
				<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}			
				Aedini	<i>Aedes</i>	<i>Stegomyia</i>	<i>scutellaris</i>	-	<i>albopictus</i> ^{d,c,z,r,j}
								-	<i>aobae</i>
								-	<i>hebrideus</i> ^d
								-	<i>pernotatus</i>
								-	<i>polynesiensis</i> ^{d,c,z,F,r}
	-	<i>lineata</i> ^r							
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>Culex</i>	<i>pipiens</i>	-	<i>pacificus</i>		
					<i>sitiens</i>	-	<i>quinquefasciatus</i> ^{f,j}		
					<i>sitiens</i>	-	<i>annulirostris</i> ^{R,J}		
					<i>sitiens</i>	-	<i>sitiens</i> ^j		
					<i>mochthogenes</i>	-	<i>femineus</i>		
					<i>fraudatrix</i>	<i>buxtoni</i>	<i>buxtoni</i>		
					-	-	<i>starckeae</i>		
					<i>Coquillettida</i>	<i>Coquillettida</i>	-	<i>xanthogaster</i> ^r	
					<i>Tripteroides</i>	<i>Polylepidomyia</i>	-	<i>folicola</i>	
					<i>Tripteroides</i>	<i>Polylepidomyia</i>	-	<i>melanesiensis</i>	
	Uranotaeniini	<i>Uranotaenia</i>	<i>Uranotaenia</i>	-	-	<i>barnesi</i>			

Legend for vector status: M or m: Malaria, D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: 39,65,121,208,257,291

Wallis and Futuna (8 species)

Subfamily	Tribe	Genus	Subgenus	Group	Complex	Species	
Culicinae	Aedini	<i>Aedes</i>	<i>Aedes</i>	<i>vexans</i>	-	<i>vexans</i> ^j	
				<i>oceanicus</i>	-	<i>oceanicus</i>	
				<i>aegypti</i>	-	<i>aegypti</i> ^{D,C,Z,r}	
				<i>scutellaris</i>	-	<i>polynesiensis</i> ^{d,c,z,F,r}	
				<i>zoromorphus</i>	-	<i>futunae</i>	
				<i>pipiens</i>	<i>pipiens</i>	<i>quinquefasciatus</i> ^j	
	Culicini	<i>Culex</i>	<i>Culex</i>	<i>Culex</i>	<i>annulirostris</i> ^{R,J}	-	<i>annulirostris</i> ^{R,J}
					<i>sitiens</i>	-	<i>sitiens</i> ^j

Legend for vector status: D or d: dengue virus, C or c: chikungunya virus, Z or z: Zika virus, F or f: lymphatic filariasis, R or r: Ross River virus, J or j: Japanese encephalitis virus. Capital letters denote primary vectors, and lower case letters denote secondary vectors.

References: 65,93,121,208,226

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